

# Introduction to Read-Based Alignment

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January 9, 2019

# Aligning to a Reference

- Aligning sequences is a classic problem
  - Early bioinformatic problem
  - Very similar to older text matching problems
- Several algorithms exist
  - Tradeoffs of speed versus accuracy, sensitivity
- Sequencing throughput creates new problems
  - Short reads have less information than long seqs
  - Data volume requires faster processing per read

# Example of alignment

Read:

TCAACTCTGCCAACACCTCCTCCAGGAAGCACTCCTGGATTCCTCTTCCAACAAGATTCTGGAGGGCA

Genome:

ATAAAATGCCAAAATTAACTAGAAGGTGAGTAGAAACTTAAATAAACTAATTACCATTGATGAGAAAAAAAATC  
TGCCACTGAAAAGGCACCCGGTCCAGAGGGTTCATGAGCGGGAACTGTAGAACCTTCGAATTCAACTCTGC  
CAACACCTCCTCCAGGAAGCACTCCTGGATTCCTCTTCCAACAAGATTCTGGAGGGCAGCTCCTCCA  
ACATGCCCAACAGCTCTGCAGACATATCATATCATATCTTCCATACCATACTGCCATGCCATACA

# Example of alignment

Read:

TCAACTCTGCCAACACCTCCTCCAGGAAGCACTCCTGGATTCCTCTGCCAACAGATTCTGGAGGGCA

Genome:

ATAAAATGCCAAAATTAACTAGAAGGTGAGTAGAAACTTAAATAAACTAATTACCATTGATGAGAAAAAAAATC  
TGCCACTGAAAAGGCACCCGGTCCAGAGGGTTCATGAGCGGGAACTGTAGAACCTTCGAATTCAACTCTGC  
CAACACCTCCTCCAGGAAGCACTCCTGGATTCCTCTGCCAACAGATTCTGGAGGGCAGCTCCTCCA  
ACATGCCCAACAGCTCTGCAGACATATCATATCATATCTTCCATACCATACTGCCATGCCATACA

# How Would You Find That?

- Brute force comparison
- Smith-Waterman
- Suffix Tree
- Burrows-Wheeler Transform

# Brute Force Method

TCGATCC  
?  
GACCTCA**TCGATCC**CACTG

# Brute Force Method

TCGATCC  
X  
GACCTCATCGATCCCACTG

# Brute Force Method

TCGATCC  
X  
GACCTCATCGATCCCACTG

# Brute Force Method

TCGATCC  
||x  
GACCTCATCGATCCCACTG

# Brute Force Method

TCGATCC  
GACCTCA~~TCGATCC~~ CACTG

# Smith-Waterman

Simplistic Scoring Scheme:

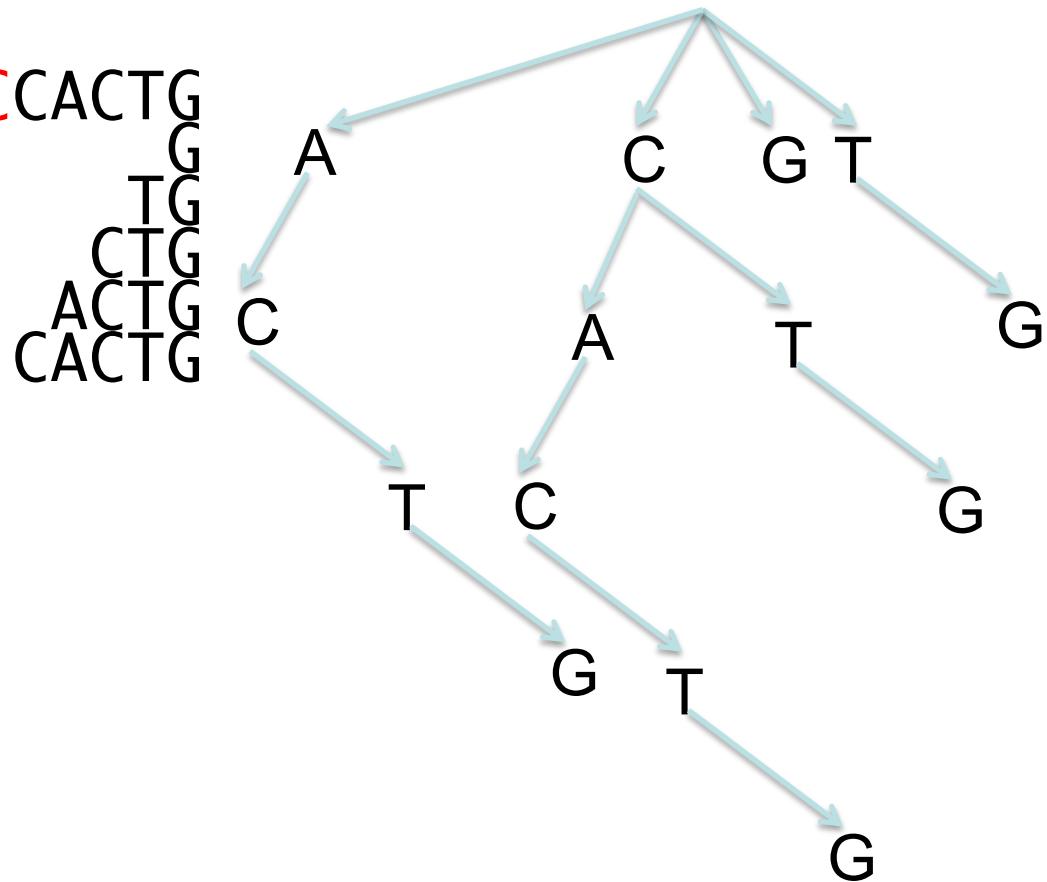
+1 match	if moving diagonally	}	take the highest of these
-1 mismatch	if moving diagonally		
-1 gap	if moving hor. or vert.		

(no penalty for terminal gaps)

	0	-1	-2	0	2	1	1	0	1	3	3	2	3	5	7	6	5	4	3	2
T	0	-1	-1	1	1	0	1	0	2	4	3	2	4	6	5	4	3	2	1	0
C	0	-1	0	0	-1	0	-1	-1	3	2	1	1	1	5	4	3	2	1	0	1
G	0	0	1	0	-1	-2	0	2	1	0	2	4	3	2	1	0	1	0	-1	-2
A	0	1	0	-1	-2	-1	1	1	0	1	3	2	1	1	1	0	-1	-2	-1	0
T	0	-1	0	-1	0	0	2	1	0	2	1	0	0	2	1	0	-1	0	0	0
C	0	-1	-1	-1	-1	1	0	-1	1	0	-1	-1	1	0	-1	-1	-1	-1	1	0
G	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
A	^	G	A	C	C	T	C	A	T	C	G	A	T	C	C	C	A	C	T	G

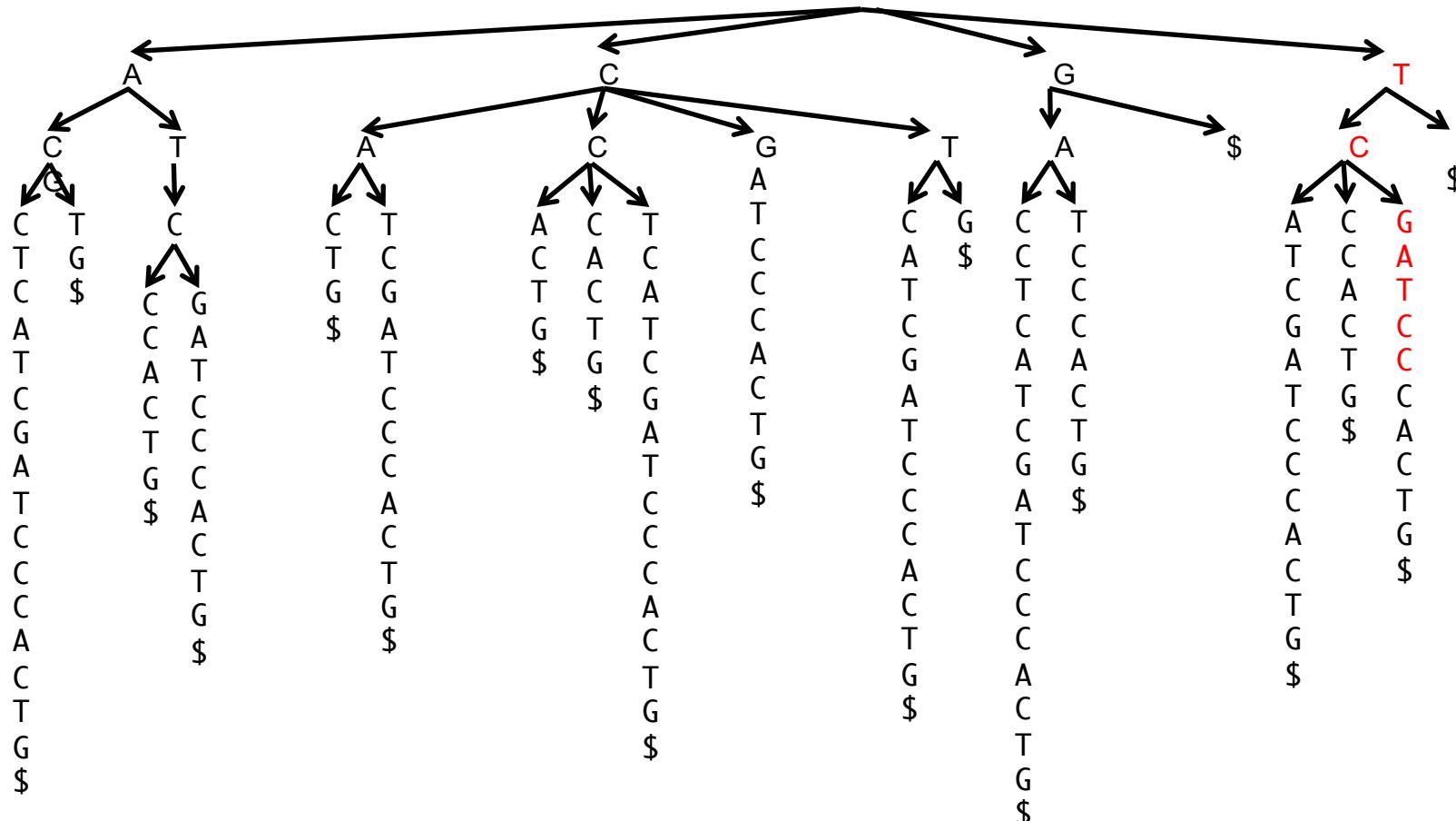
# Suffix Tree

GACCTCA**TCGATCC**CACTG



# Suffix Tree

GACCTCA**TCGATCC**ACTG



# Burrows-Wheeler Transform

GACCTCA~~TCGATCC~~CACTG\$  
ACCTCA~~TCGATCC~~CACTG\$G  
CCTCA~~TCGATCC~~CACTG\$GA  
CTCA~~TCGATCC~~CACTG\$GAC  
TCA~~TCGATCC~~CACTG\$GACC  
CATCGATCCCACTG\$GACCT  
~~ATCGATCC~~CACTG\$GACCTC  
TCGATCCCACTG\$GACCTCA  
CGATCCCAGTG\$GACCTCA~~T~~  
GATCCCACTG\$GACCTCA~~T~~  
~~ATCC~~CACTG\$GACCTCA~~TCG~~  
TCCCACTG\$GACCTCA~~TCGA~~  
CCCACTG\$GACCTCA~~TCGAT~~  
CCACTG\$GACCTCA~~TCGATC~~  
CACTG\$GACCTCA~~TCGATCC~~  
ACTG\$GACCTCA~~TCGATCC~~  
CTG\$GACCTCA~~TCGATCCA~~  
TG\$GACCTCA~~TCGATCC~~CAC  
G\$GACCTCA~~TCGATCC~~CACT  
\$GACCTCA~~TCGATCC~~CACTG

ACCTCA~~TCGATCC~~CACTG\$G  
ACTG\$GACCTCA~~TCGATCC~~  
~~ATCC~~CACTG\$GACCTCA~~TCG~~  
~~ATCGATCC~~CACTG\$GACCTC  
CACTG\$GACCTCA~~TCGATCC~~  
CATCGATCCCACTG\$GACCT  
CCACTG\$GACCTCA~~TCGATC~~  
CCCACTG\$GACCTCA~~TCGAT~~  
CCTCA~~TCGATCC~~CACTG\$GA  
CGATCCCAGTG\$GACCTCA~~T~~  
CTCATCGATCCCACTG\$GAC  
CTG\$GACCTCA~~TCGATCCA~~  
GACCTCA~~TCGATCC~~CACTG\$  
GATCCCACTG\$GACCTCA~~T~~  
G\$GACCTCA~~TCGATCC~~CACT  
TCATCGATCCCACTG\$GACCT  
TCCCACTG\$GACCTCA~~TCGA~~  
TCGATCCCACTG\$GACCTCA  
TG\$GACCTCA~~TCGATCC~~CAC  
\$GACCTCA~~TCGATCC~~CACTG



# How Do We Use This To Align?

GAC	
CAC	
GAT	
CAT	
CCA	
→ TCA	
CCC	
→ TCC	
ACC	
→ TCG	
CCT	
ACT	
\$GA	
CGA	
→ TG\$	
CTC	
ATC	
ATC	
CTG	
G\$G	

- Start with the transform column
- My read starts with a T, so I want rows with Ts in them
- This column gives me all the single nucleotide counts
- Sort the single nucleotide counts to get the alphabetically first column
- Now these two columns give me all the dinucleotide counts
- Sort those to get the alphabetically first two columns
- Now there is only one place my read can match

# FM Index

G  
G  
G  
C  
C  
T  
C  
T  
A  
T  
C  
A  
\$  
C  
T  
C  
A  
A  
C  
G

- Start with the transform column
- Count all the characters, sort them, and store the count of lower characters

A	C	G	T	\$
0	4	12	15	19

- This gives the positions of all the bases in the first column (because it's sorted)

# FM Index

A  
A  
A  
A  
C  
C  
C  
C  
C  
T  
C  
T  
A  
T  
C  
A  
\$  
C  
T  
C  
A  
A  
C  
G

- Take the query sequence **TCGATCC**
- Start at the end and use the count table to look up the position of the *last base* in the *first column*

A	C	G	T	\$
0	4	12	15	19

- The last column comes immediately before the first column
- Find all the rows of the last column with the next to last base

# FM Index

AC  
AC  
AT  
AT  
CA  
CA  
**CC**  
**CC**  
**CC**  
CG  
CT  
CT  
GA  
GA  
G\$  
TC  
TC  
TC  
TG  
\$G

G  
G  
G  
C  
C  
**T**  
**C**  
**T**  
A  
T  
C  
A  
\$  
C  
T  
C  
A  
A  
C  
G

- Take the query sequence **TCGATCC**
- The order of a given character in the last column matches the order of the same instance of that character in the first column
- The 3<sup>rd</sup>-5<sup>th</sup> Cs in the last column precede Cs in the first column, so we now want the 3<sup>rd</sup>-5<sup>th</sup> Cs in column 1

A	C	G	T	\$
0	4	12	15	19

- Now we take the next character and look for Ts in the last column (the 2<sup>nd</sup> T)

# FM Index

ACC  
ACT  
ATC  
ATC  
CAC  
CAT  
CCA  
CCC  
CCT  
CGA  
CTC  
CTG  
GAC  
GAT  
G\$G  
TCA  
**TCC**  
TCG  
TG\$  
\$GA

G  
G  
G  
C  
C  
T  
C  
T  
A  
T  
C  
A  
\$  
C  
T  
C  
A  
A  
C  
G

- Take the query sequence **TCGATCC**
- The second T is preceded by the 3<sup>rd</sup> A

A	C	G	T	\$
0	4	12	15	19

# FM Index

ACCT  
ACTG  
**ATCC**  
ATCG  
CACT  
CATC  
CCAC  
CCCA  
CCTC  
CGAT  
CTCA  
CTG\$  
GACC  
GATC  
G\$GA  
TCAT  
TCCC  
TCGA  
TG\$G  
\$GAC

**G**  
G  
**G**  
C  
C  
T  
C  
T  
A  
T  
C  
A  
\$  
C  
T  
C  
A  
A  
C  
G

- Take the query sequence **TCGATCC**
- The third A is preceded by the 2<sup>nd</sup> G

A	C	G	T	\$
0	4	12	15	19

# FM Index

ACCTC  
ACTG\$  
ATCCC  
ATCGA  
CACTG  
CATCG  
CCACT  
CCCAC  
CCTCA  
CGATC  
CTCAT  
CTG\$G  
GACCT  
**GATCC**  
G\$GAC  
TCATC  
TCCA  
TCGAT  
TG\$GA  
\$GACC

G  
C  
G  
C  
C  
T  
C  
T  
A  
T  
C  
A  
\$  
C  
T  
C  
A  
A  
C  
G

- Take the query sequence **TCGATCC**
- The second G is preceded by the 6<sup>th</sup> C

A	C	G	T	\$
0	4	12	15	19

# FM Index

ACCTCA  
ACTG\$G  
ATCCCCA  
ATCGAT  
CACTG\$  
CATCGA  
CCACTG  
CCCACT  
CCTCAT  
**CGATCC**  
CTCATC  
CTG\$GA  
GACCTC  
GATCCC  
G\$GACC  
TCATCG  
TCCCAC  
TCGATC  
TG\$GAC  
\$GACCT

G  
G  
G  
C  
C  
C  
T  
C  
T  
A  
T  
C  
A  
\$  
C  
T  
C  
A  
A  
C  
G

- Take the query sequence **TCGATCC**
- The sixth C is preceded by the 3<sup>rd</sup> T

A	C	G	T	\$
0	4	12	15	19

# FM Index

ACCTCAT  
ACTG\$GA  
ATCCCAC  
ATCGATC  
CACTG\$G  
CATCGAT  
CCACTG\$  
CCCACTG  
CCTCATC  
CGATCCC  
CTCATCG  
CTG\$GAC  
GACCTCA  
GATCCA  
G\$GACCT  
TCATCGA  
TCCCACT  
**TCGATCC**  
TG\$GACC  
\$GACCTC

G  
G  
G  
C  
C  
C  
T  
C  
T  
A  
T  
C  
A  
\$  
C  
T  
C  
A  
A  
C  
G

- Take the query sequence **TCGATCC**
- And we're done

A	C	G	T	\$
0	4	12	15	19

- To find the position in the genome, we keep a separate index of positions for a sparse set of rows in the table and then just walk through the transform to the nearest indexed row

# Heuristic Improvements

- Seeding
  - Use a hash of exact matches to limit searches
  - Use varying hash types
  - Seed with suffix tree/array or B-W then extend
- Limiting mismatches or indels
  - Constrain poor quality searches
  - Known as ‘banded’ in Smith-Waterman
- Using base quality to guide backtracking

# Common Short Read Aligners

- Seed and Smith-Waterman extend
  - Novoalign
- BWA align gap-free
  - Bowtie
- BWA align with gaps
  - BWA aln, Bowtie2
- BWA Seed and Smith-Waterman extend
  - BWA mem
- Seed clustering and stitching
  - STAR, Blasr, minimap2