

Evolution and genomics, Cesky Krumlov

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# Genomic analyses using RADseq:

## 1. Raw data manipulation

# Demo

Upload and inspection of a raw Illumina sequence data set (stickleback RAD sequences from the Misty system, Canada, unpublished; 100k subset from a single SE100 Illumina lane)

```
library(ShortRead)
# d<-readFastq(dirPath='C:/Users/daniel/Documents/science/teaching/cesky
# krumlov 2016/course.materials/R.files',
# pattern='illumina.SE100.fastq', withIds=T)
d

## class: ShortReadQ
## length: 100000 reads; width: 100 cycles

# Call the read IDs of the ShortRead object
id(d)[1:4] # just the first four elements

## A BStringSet instance of length 4
## width seq
## [1] 53 BS-DSFCNTROL03:312:C3...1101:1232:2073 1:Y:0:
## [2] 53 BS-DSFCNTROL03:312:C3...1101:1213:2079 1:Y:0:
## [3] 53 BS-DSFCNTROL03:312:C3...1101:1185:2091 1:Y:0:
## [4] 53 BS-DSFCNTROL03:312:C3...1101:1102:2093 1:Y:0:
```

```

# Call the sequences
sread(d)[1:5]

## A DNAStringSet instance of length 5
## width seq
## [1] 100 CTGAATGCAGGTCATTTTGGTN...CGGACGNNNTCCCTGCATCCT
## [2] 100 TAGCATGCAGGAAGTCCGTTGN...CAACTCNNNAAAATTTGCCAA
## [3] 100 GCGCCTGCAGGGCTTTATCCAG...GCTGCTGNNCAGATGTCCTCC
## [4] 100 AGGTATATGCACAAAATGAGAT...CAATCTTNNCAAGCAACAGCA
## [5] 100 NCACGTGCACAAAAAAGAGT...NNNNNNNNNNNNNNNNNNNNNN

# ... and their qualities
quality(d)[1:5]

## class: FastqQuality
## quality:
## A BStringSet instance of length 5
## width seq
## [1] 100 ;;<;@@2<@2222;@<<><;=#...#####
## [2] 100 <<<??@<@=222@00??@??#...#####
## [3] 100 <<<@002<@22<@000?@?@00...#####
## [4] 100 <<<>?26@=)9>?@0000?<?...#####
## [5] 100 #07>@222:))2=>@?@<;9>=...#####

```

## Pattern matching, counting

```
grep("TATATATATATATATATATA", sread(d))
```

```
## [1] 6053 15441 82461 84384
```

```
match <- grep("TATATATATATATATATATA", sread(d))  
length(match)
```

```
## [1] 4
```

## Subsetting of ShortRead object

```
d.match <- d[match]  
sread(d.match)
```

```
## A DNASTringSet instance of length 4  
## width seq  
## [1] 100 TAGCATGCAGGGAGGCCTGTGT...TATTTTACACACAACGACAGA  
## [2] 100 CTAGGTGCAGGTACAGTGATCG...TGCCTGCTCCCGACCGGCTTC  
## [3] 100 CTGATGCAGGACAGGTCCTCCC...ATATATATATATATATATATC  
## [4] 100 TAATGTGCAGGAGTCTGTAGTC...TATATATATATATATATATAT
```

## Cleaning a ShortRead object

```
d.clean <- clean(d) # remove all reads with >= 1 'N'  
sread(d.clean)[1]  
  
## A DNASTringSet instance of length 1  
## width seq  
## [1] 100 CCATGTTGCAGGTGTGAAGGCT...GGGGACACGCCGGCCGTTTGC
```

## Trimming a ShortRead object

```
d.trim <- narrow(d.match, start = 1, end = 10) # either end or width  
quality(d.trim)  
  
## class: FastqQuality  
## quality:  
## A BStringSet instance of length 4  
## width seq  
## [1] 10 ==>A<224?2  
## [2] 10 BBCDF224A2  
## [3] 10 @@@FFADDA2  
## [4] 10 CCCFF222C2
```

## Write a ShortRead object out as fastq file

```
# writeFastq(d.clean,  
# file='C:/Users/daniel/Documents/science/teaching/cesky  
# krumlov  
# 2016/course.materials/R.files/my.clean.reads.fastq')
```

# Tasks

- ▶ Upload the stickleback data set *illumina.SE100.fastq*
- ▶ Inspect the ID, sequence and quality of the reads 1000 to 1002
- ▶ Generate a new object X containing the data from the reads 10001-20000
- ▶ Determine the proportion of X's reads containing one or more 'N', and eliminate them from X
- ▶ What proportion of the filtered X is derived from the individual with barcode (first five bases) 'CGATA'?
- ▶ Derive the object Y from X, including only these specific CGATA-reads. Confirm that this worked by inspecting the reads
- ▶ What proportion of Y's reads contains the correct restriction enzyme overhang 'TGCAGG' at the correct position (i.e., following the barcode)? Copy these reads to object Z
- ▶ Clip the barcodes from Z, then write Z out as a fastq file