

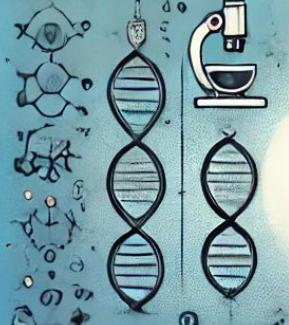
# ALIGNMENT

Rayan Chikhi  
Institut Pasteur

RAYAN CHIKHI

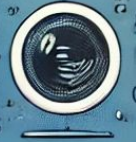
RAYAN CHIKHI

INSTITUT PASTEUR



# ALIGNMENT

RAYAN CHIKHI



# Hello!

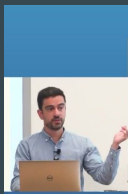
- I'm a researcher in bioinformatics algorithms
- *de novo* assembly, big data, some alignment, k-mers, pangenomics. Week 1 stuff :)

 @RayanChikhi on X/Bsky

<http://rayan.chikhi.name>

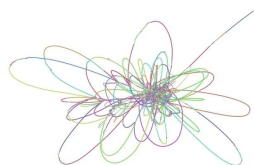
<https://github.com/IndexThePlanet/Logan>

Sequence  
Bioinformatics



Rayan Chikhi

k=127



# Course objectives

- **Enough background** to understand the alignment methods in an article
- Increase confidence in using alignment tools
- Understand **why** alignment is not so straightforward actually

# Course outline

- **Fundamentals**
- Many **flavors** and **tools** for pairwise DNA alignment
- **m-multiple**  
sequence  
alignment
- Alignment to **databases**
- Into the unknown: profile and structure search

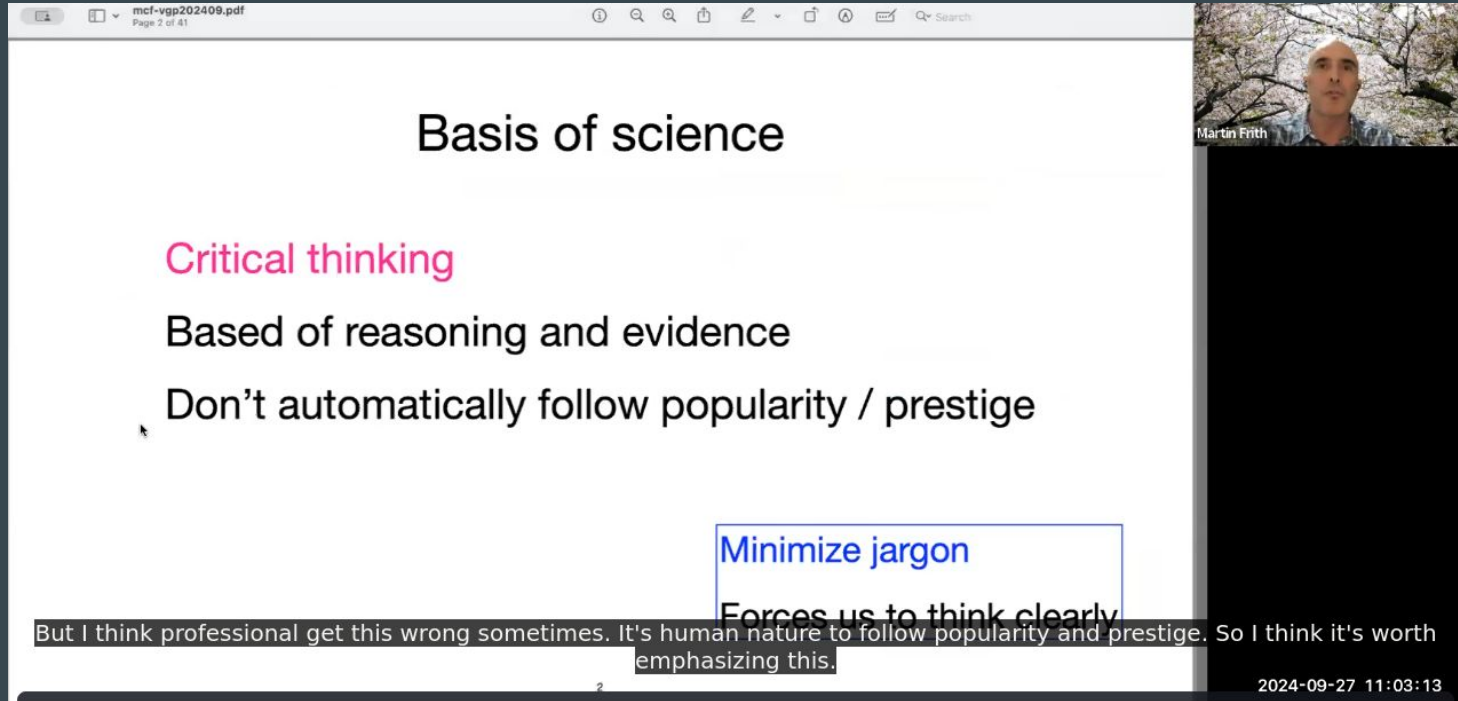
## Course diff, if you've seen it

- **part 1:** theory refined, follows last year mostly
- **part 2:** more applied, demos

# Shoot-out to inspirations: Mike Zody's previous lectures



# Shoot-out to inspirations: Martin Frith's talk and papers



mcf-vgp202409.pdf  
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## Basis of science

Critical thinking

Based on reasoning and evidence

Don't automatically follow popularity / prestige

Minimize jargon

Forces us to think clearly

But I think professionals get this wrong sometimes. It's human nature to follow popularity and prestige. So I think it's worth emphasizing this.

Martin Frith

2024-09-27 11:03:13

(Stopping on this slide for a second because wow)



Shoot-out to inspirations: Milos, Joan 🥲, workshop team, workshop participants: all who provided feedback last year (& RC Edgar)



# Questions to the audience

1. Have you ever **run** a sequence alignment software?
2. Was it **directly** or as part of a pipeline?
3. Done **multiple** sequence alignment?
4. Know who/what **Smith-Waterman** is?

# What's an “alignment”?

```
...C   A   T   A   G...  
...C   G   T   -   G...  
...match mismatch match deletion match...
```

Given two (or more) sequences, determine how the letters best **line up**, to capture **evolutionary relationships**.

# The many types of alignments

Pairwise (2 sequences)

Score	Expect	Identities	Gaps	Strand
435 bits(235)	5e-117	360/410(88%)	50/410(12%)	Plus/Minus
Query 302	GTAGGACAGGTGCCGGCAGCGCTCTGGGTCATTTTCGGCGAGGACCGCTTTCGCTGGAG-			360
Sbjct 3589	GTAGGACAGGTGCCGGCAGCGCTCTGGGTCATTTTCGGCGAGGACCGCTTTCGCTGGAGC			3531
Query 361	-----ATCGGCCTGTCGCTTGCGGTATTCGGAATCTTGACGCCCTCGCTCAAGCC			411
Sbjct 3529	GCGACGATGATCGGCCTGTCGCTTGCGGTATTCGGAATCTTGACGCCCTCGCTCAAGCC			3471

<https://techcommunity.microsoft.com/t5/azure-high-performance-computing/running-ncbi-blast-on-azure-performance-scalability-and-best-practices/p2410483>

# The many types of alignments

## Multiple sequences (>2)

Q5E940_BOVIN	-----MPREDRATWKSNYFLKIIQLDDVYKCFIVGADNVGSKOMQIIRMSLRGK-AVVLGMKNTMMRKAIRGHLENN--PALE	76
RLA0_HUMAN	-----MPREDRATWKSNYFLKIIQLDDVYKCFIVGADNVGSKOMQIIRMSLRGK-AVVLGMKNTMMRKAIRGHLENN--PALE	76
RLA0_MOUSE	-----MPREDRATWKSNYFLKIIQLDDVYKCFIVGADNVGSKOMQIIRMSLRGK-AVVLGMKNTMMRKAIRGHLENN--PALE	76
RLA0_RAT	-----MPREDRATWKSNYFLKIIQLDDVYKCFIVGADNVGSKOMQIIRMSLRGK-AVVLGMKNTMMRKAIRGHLENN--PALE	76
RLA0_CHICK	-----MPREDRATWKSNYFMKIIQLDDVYKCFIVGADNVGSKOMQIIRMSLRGK-AVVLGMKNTMMRKAIRGHLENN--PALE	76
RLA0_RANSY	-----MPREDRATWKSNYFLKIIQLDDVYKCFIVGADNVGSKOMQIIRMSLRGK-AVVLGMKNTMMRKAIRGHLENN--SALE	76
Q7ZUG3_BRARE	-----MPREDRATWKSNYFLKIIQLDDVYKCFIVGADNVGSKOMQIIRMSLRGK-AVVLGMKNTMMRKAIRGHLENN--PALE	76
RLA0_ICTPU	-----MPREDRATWKSNYFLKIIQLNDVYKCFIVGADNVGSKOMQIIRMSLRGK-AVVLGMKNTMMRKAIRGHLENN--PALE	76
RLA0_DROME	-----MVRENKAAWKAQYFIKVVLELDFEYKCFIVGADNVGSKOMQIIRMSLRGK-AVVLGMKNTMMRKAIRGHLENN--PALE	76
RLA0_DICDI	-----MSAGC-SKRKKLFLKATKLFITTDKMIVAEADYVGSGLQKIRKRSIRGI-GAVLMGKNTMIRKVIIRDADSK--PELD	75
Q54LP0_DICDI	-----MSAGC-SKRKNVFLKATKLFITTDKMIVAEADYVGSGLQKIRKRSIRGI-GAVLMGKNTMIRKVIIRDADSK--PELD	75
RLA0_PLAFB	-----MAKLSKQKQKQMYLEKLSLISLQQYSKILIVHVDNVGSKOMQIIRMSLRGK-AVVLGMKNTMMRKAIRGHLENN--PALE	76
RLA0_SULAC	-----MIGLAVTTTCKIAKWKVDEVAELIKLTKHTILIANIEGFPADKLHEIRKRLRGK-ADIKVTKNNLFNIALKNAG----YDTP	79
RLA0_SULTO	-----MRIMAVITQERIAKWKIEEYKLEKQKLERYHTILIANIEGFPADKLHDIRKRMGM-AEIKVTKNTLFGIAAKNAG----LDVS	80
RLA0_SULSO	-----MKRLALALQKQKVASWKEEYKELTELKIKNSMTLIGNLEGFPADKLHEIRKRLRGK-ATIKVTKNTLFGIAAKNAG----IDIP	80
RLA0_AERPE	MSVVSIVGQMYKREKPTPEWKTLMLERELELFSKIRVVLFDLTCPTFFVYQVYVKKLWKK-YPMVVAKKRIILRAMKAAGLE--LDDN	86
RLA0_PYRAE	-----MMLAIGKRRYVTRQYDARKVKIYSEATELLOKQYVYVFLDGLGSLRILHEVYRRLRY-GVTKIIPLEKIAFTKVVYGG--IPAE	85
RLA0_METAC	-----MAEERHTEHTPQWKDEIENIKELIQSHKVFQMGVIEGLLATKMKQKIRRDLDV-AVLKVSRTNLEERALNQLG----ETIP	78
RLA0_METMA	-----MAEERHTEHTPQWKDEIENIKELIQSHKVFQMGVRIEGLLATKMKQKIRRDLDV-AVLKVSRTNLEERALNQLG----ESIP	78
RLA0_ARCFU	-----MAAVRGS--PPEYKVRAVEETKRMISKPVVAIVSFENVPAGOMCKIRREFRGK-AEIKVVKNTLLEERALDGLG----GDYL	75
RLA0_METKA	MAVKAKQPPSGYEYKVAEKKRREYKELKELMDVENYGLVDLEGIYAPOLQEIIRAKLREERDILIRMSRNTLMRILALEEKLDER--PELE	78
RLA0_METH	-----MAHVAEWKKKEVEELANLKSYPVIALVDVSSMAYPLSQMRRLLIRENGLLRVSRNTLIELAIKKAQELGKPELE	84
RLA0_METT	-----MITAESEHKIAPWKIEEYVKLKEELKNGQIIVALVDMMEVPARLOQEIIRDKIR-GTMTLKMSRNTLIEERAIKVAEETGNPEFA	82
RLA0_METVA	-----MIDAKSEHKIAPWKIEEYVKLKEELKNSANVIALDMMEVPARLOQEIIRDKIR-DQMTLKMSRNTLIEERAIKVAEETGNPEFA	82
RLA0_METJA	-----METKVKAHVAWKIEEYVKLKEELKSKPVAIVDMMDVYAPOLQEIIRDKIR-DKVKLRMSRNTLIEERAIKVAEELNPKLA	81
RLA0_PYRAB	-----MAHVAEWKKKEVEELANLKSYPVIALVDVSSMAYPLSQMRRLLIRENGLLRVSRNTLIEERAIKKAQELGKPELE	77
RLA0_PYRHO	-----MAHVAEWKKKEVEELANLKSYPVIALVDVSSMAYPLSQMRRLLIRENGLLRVSRNTLIEERAIKKAQELGKPELE	77
RLA0_PYRKO	-----MAHVAEWKKKEVEELANLKSYPVIALVDVAGVYAPYPLSKMRDKLR-GKALLRVSRNTLIEERAIKKAQELGKPELE	76
RLA0_HALMA	MSAESERTETPEWQEEYDAIVMIESVESVGVNIAAGIYRGLQDMRRDLHGT-AELRVSRNTLIEERALDDVD--DGLF	79
RLA0_HALVO	MSSEVQRTEVTPQWKREEVDELVDYIESVESVGVVAGIYRGLQDMRRDLHGT-AELRVSRNTLIEERALDDVD--DGLF	79
RLA0_HALSA	MSAEQRTTEVPEWKRQEVDELVDYIESVESVGVVAGIYRGLQDMRRDLHGT-AALRVSRNTLIEERALEEAG--DGLD	79
RLA0_THEAC	-----MKEVSQKKELVNETDTRIKASRSVAIVDAGIRTRQIDDIRKKNRGK-INLVKIKKTLLEKALENLGD--EKLS	72
RLA0_THEVO	-----MRKINPKKKEIVSELAQDITKSKAVAVDVKIGRTRQIDDIRKKNRDK-VKIKVVKKTLLEKALDIND--EKLT	72
RLA0_PICTO	-----MTEFAQKIDFVKNLENEINSRKRVAIVISIKGLRNNFQKIKRNSIRDK-ARIKVSRRARLLRLALENTGK--NNIV	72
ruler	1 . . . . . 10 . . . . . 20 . . . . . 30 . . . . . 40 . . . . . 50 . . . . . 60 . . . . . 70 . . . . . 80 . . . . . 90	

[https://en.wikipedia.org/wiki/Multiple\\_sequence\\_alignment](https://en.wikipedia.org/wiki/Multiple_sequence_alignment)

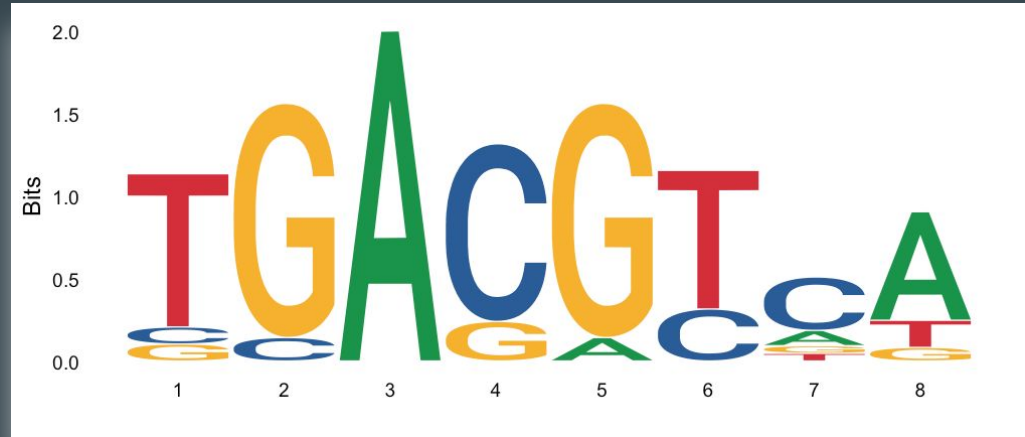
# The many types of alignments

1 sequence versus a database

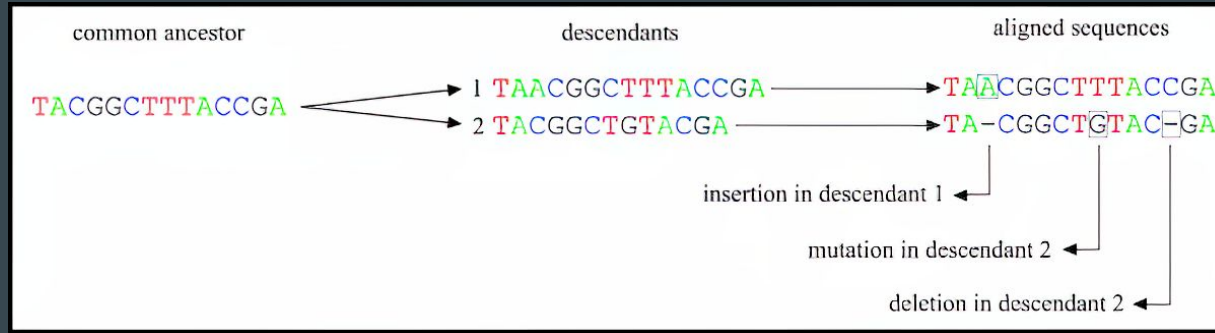
Sequences producing significant alignments:		Score	E
		(bits)	Value
<a href="#">sp P14120 RL30</a>	YEAST 60S RIBOSOMAL PROTEIN L30 (YL32) (RP73)	201	3e-52
<a href="#">sp P38664 RL30</a>	KLULA 60S RIBOSOMAL PROTEIN L30 (L32)	190	7e-49
<a href="#">sp P52808 RL30</a>	SCHPO 60S RIBOSOMAL PROTEIN L30 (L32)	137	5e-33
<a href="#">sp P04645 RL30</a>	HUMAN 60S RIBOSOMAL PROTEIN L30	129	1e-30
<a href="#">sp O49884 RL30</a>	LUPLU 60S RIBOSOMAL PROTEIN L30	129	1e-30
<a href="#">sp P47833 RL30</a>	CHICK 60S RIBOSOMAL PROTEIN L30	129	1e-30
<a href="#">sp O48558 RL30</a>	MAIZE 60S RIBOSOMAL PROTEIN L30	127	4e-30
<a href="#">sp P49153 RL30</a>	TRYBB 60S RIBOSOMAL PROTEIN L30	113	9e-26
<a href="#">sp P39095 RL30</a>	LEIMA 60S RIBOSOMAL PROTEIN L30	108	2e-24
<a href="#">sp O27127 RL3E</a>	METTH 50S RIBOSOMAL PROTEIN L30E	79	2e-15
<a href="#">sp O74018 RL3E</a>	PYRHO 50S RIBOSOMAL PROTEIN L30E	71	5e-13
<a href="#">sp Q9YAU3 RL3E</a>	AERPE 50S RIBOSOMAL PROTEIN L30E	71	5e-13
<a href="#">sp P14025 RL3E</a>	METVA 50S RIBOSOMAL PROTEIN L30E	69	2e-12
<a href="#">sp P54061 RL3E</a>	METJA 50S RIBOSOMAL PROTEIN L30E	66	1e-11
<a href="#">sp P11522 RL3E</a>	SULAC 50S RIBOSOMAL PROTEIN L30E (ORF 104)	64	9e-11
<a href="#">sp P29160 RL3E</a>	THECE 50S RIBOSOMAL PROTEIN L30E	64	9e-11
<a href="#">sp O28389 RL3E</a>	ARCFU 50S RIBOSOMAL PROTEIN L30E	60	1e-09
<a href="#">sp O59165 RS6X</a>	PYRHO 30S RIBOSOMAL PROTEIN HS6-LIKE	37	0.011
<a href="#">sp P54066 RS6X</a>	METJA 30S RIBOSOMAL PROTEIN HS6-LIKE	35	0.043
<a href="#">sp O26355 RS6X</a>	METTH 30S RIBOSOMAL PROTEIN HS6-LIKE	34	0.056
<a href="#">sp P55858 RS6X</a>	SULSO 30S RIBOSOMAL PROTEIN HS6-LIKE	34	0.056
<a href="#">sp Q9YAX7 RS6X</a>	AERPE 30S RIBOSOMAL PROTEIN HS6-LIKE	33	0.17
<a href="#">sp P34667 YO11</a>	CAEEL HYPOTHETICAL 20.8 KD PROTEIN ZK686.1 I...	33	0.17
<a href="#">sp O29494 RS6X</a>	ARCFU 30S RIBOSOMAL PROTEIN HS6-LIKE	32	0.28
<a href="#">sp Q55681 RECG</a>	SYNY3 ATP-DEPENDENT DNA HELICASE RECG	32	0.37
<a href="#">sp P46055 ERF1</a>	HUMAN EUKARYOTIC PEPTIDE CHAIN RELEASE FACTO...	30	1.1
<a href="#">sp P35615 ERF1</a>	XENLA EUKARYOTIC PEPTIDE CHAIN RELEASE FACTO...	29	2.5
<a href="#">sp P52291 PPA1</a>	PICPA ACID PHOSPHATASE PHO1 PRECURSOR	29	3.2
<a href="#">sp P55768 YLXQ</a>	ENTFC PROBABLE RIBOSOMAL PROTEIN IN INF6 5'R...	28	4.2
<a href="#">sp Q01056 TEGU</a>	HSVSA PROBABLE LARGE TEGUMENT PROTEIN	28	4.2
<a href="#">sp P55124 LKTC</a>	PASSP LEUKOTOXIN-ACTIVATING LYSINE-ACYLTRANS...	27	7.3
<a href="#">sp O25074 Y303</a>	HELPHY PROBABLE GTP-BINDING PROTEIN HPO303	27	9.5

# The many types of alignments

1 sequence versus a profile



# Why align?



<https://users.ugent.be/~avierstr/principles/aligning.html>

One of the two pillars of sequence bioinformatics (with assembly).

Variant calling, RNA-seq quantification, taxonomic classification, etc..



# How to do molecular biology



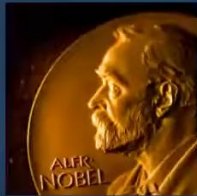
1. Sequences



2. Alignment



3. Tree, structure, function...



4. Publish

# What can be aligned? Many things..:

DNA vs DNA

RNA vs RNA

DNA vs RNA,

Protein sequence vs DNA,

Protein sequence vs protein sequence,

Protein structure vs protein structure,

etc..

# Some vocabulary

**Query**: sequence to align

**Reference** (or **target**): other sequence to align to

**Hit** (or **match** or **alignment**): part of query aligned to part of reference

**Homology**: *shared ancestry*

**Similarity**, **identity**: mathematical ways to detect homology

**String**: sequence

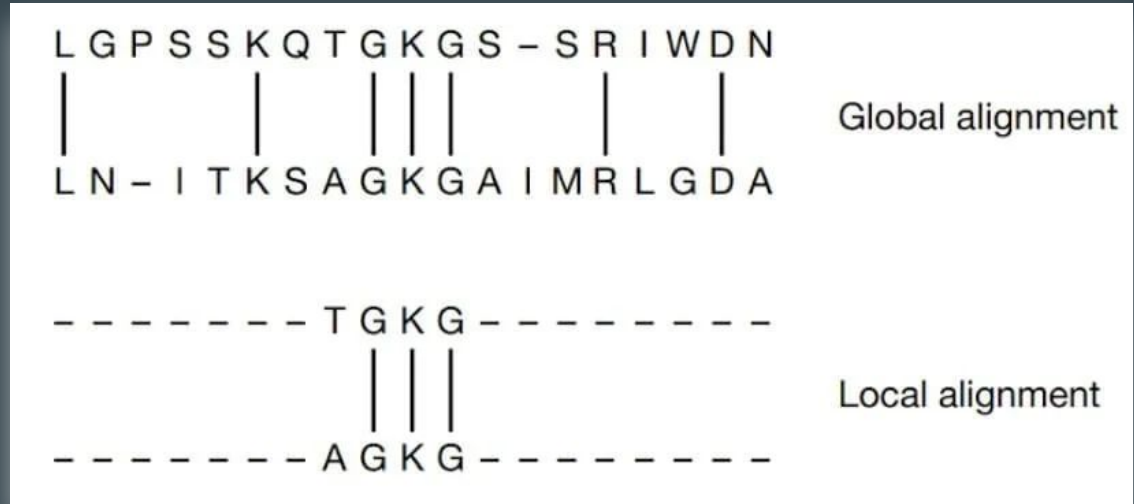
**Letter** (or **residue** or **monomer**): base pair or nucleotide or amino-acid

# Pairwise DNA

General techniques



# Global vs local



<https://microbenotes.com/local-global-multiple-sequence-alignment/>

**Global**: must align **all** nucleotides, using insertions/deletions if necessary

**Local**: you're allowed to skip beginning and/or end of either sequence

# Alignment is based on scoring

What is a *good* alignment?

One that **minimizes** a **penalty** (or **maximizes** a score).

E.g. here a mismatch gives 1 penalty, a deletion gives 2 penalties:

r: T**A**C

r: G**A**T

q: T**T**C

q: G-T

penalty=1

penalty=2

**Example: (global alignment)** here a mismatch gives 1 penalty, a deletion gives 2 penalties.

r : CAAGTTA

q : CAT-GGA

MMXDXXM

total penalty: 5

Is it the best we can do?

can also be  
aligned as:

CAAGTTA

CATG-GA

MMXMDXM

total penalty: 4

**better!**

# CIGAR strings (“Concise Idiosyncratic Gapped Alignment Report”)

*A succession of M,X,I,D letters to represent an alignment.*

M = match

I = insertion (gap in the **target** sequence)

X = mismatch

D = deletion (gap in the **query** sequence)

\* some programs use M for both matches and mismatches  $\backslash\_(\s)\_f$ , others use = instead of M

r : CAAGTTA

q : CAT-GGA

MMXDXXM (also written 2M1X1D2X1M), means: “to align the **query** to the **target**, do 2 matches, 1 mismatch, 1 deletion, 2 mismatches, 1 match”



# Exercise 1

Write the CIGAR string for this alignment:

target: GATCA-TGA

query: G-CAACCA-

Recall:

M = match

X = mismatch

I = insertion (gap in the **target** sequence)

D = deletion (gap in the **query** sequence)

# Solution

Write the CIGAR string for this alignment:

target: GATCA-TGA

query: G-CAACCA-

MDXXMIXXD

Quite high penalty alignment. It's unlikely any tool would output it. Those sequences are probably not evolutionarily related.

# Is it possible to know the lowest possible penalty for aligning 2 seqs?

i.e. the “best” alignment according to score

-> Yes! but you have to pay a price

(The price is a rather complex algorithm, that we'll see next, and the risk that the alignment isn't relevant)



 @CethanLeahy

Me: oh wow, this shop has everything my heart desires!  
Spooky shopkeeper: yes, I will warn you... every item comes with a price.  
Me: yes, I know how shops work

---

 kittydesade

Spooky Shopkeeper: The price may be more than you expect to pay.  
Me: Yes, I know how US taxes work, too.

---

 del3141

Shopkeeper, increasingly exasperated: I'm trying to tell you that I'm evil and offering these wares with no regard for the harm they will do!  
Me, also increasingly exasperated: I know what capitalism is too goddammit

# A special case: only mismatches

**Hamming** (= Manhattan) distance, A and B sequences of same length:

*Minimum number of substitutions to turn sequence A into sequence B*

e.g.

ACTAGATG

CGTACATG

# A special case: only mismatches

**Hamming** (= Manhattan) distance, A and B sequences of same length:

*Minimum number of substitutions to turn sequence A into sequence B*

e.g.

ACTAGATG

Hamming distance: 3

CGTACATG

Quick to calculate, just walk along both strings

# A harder case: mismatches and indels

How to find **lowest penalty alignment** with **mismatches AND indels**?

*(Can we still scan the seqs from left to right and decide on the fly?)*

To see this, consider aligning:    r : ACAG

  q : AGACTG

Novice level:

ACAG--

AGACTG

penalty=2 X's and 2 I's

Expert level:

Hint: gaps elsewhere

(there are other solutions)

## Exercise 2

Find a good (=low penalty) global alignment for these two sequences:

ref: ACTAGATG

query: GTACAT

Give the CIGAR string

Given that:

a mismatch (X) has 1 penalty,  
a deletion (D) has 2 penalty,  
a match (M) has no penalty  
hint: no insertions

# Solution

Find a good (=low penalty) global alignment for these two sequences:

ACTAGATG

-GTACAT-

**DXMMXMMD**

**total penalty = 6**

a mismatch (X) has 1 penalty,  
a deletion (D) has 2 penalty,  
a match (M) has no penalty



# Exercice

Just as a note, the best local alignment is:

ACTAGATG

GTACAT

XMMXMM

total penalty = 2

a mismatch (X) has 1 penalty,  
a deletion (D) has 2 penalty,  
a match (M) has no penalty

# Penalties / scores

So far we've used penalties:

- a mismatch (X) has 1 penalty,
- a deletion (D) has 2 penalty,
- a match (M) has no penalty

We will now switch to scores:

- a mismatch (X) has -1 score,
- a deletion (D) has -2 scores,
- a match (M) has +1 score

# Finding best alignments

Think about CIGAR strings, and imagine you are ChatGPT.

Somebody gave you

```
CATATGATGACAC  
CAGAGGGAATGCT
```

to align.

How would you like ChatGPT to respond?

- take a deep breath
- think step by step
- i have no fingers
- i will tip \$200
- do it right and i'll give you a nice doggy treat

You output the CIGAR letters **one by one** . So far you've said:

```
MMXXMIMMIMMDMX
```

You are GPT5 so this is indeed the **beginning** of the **best alignment** :

```
CATAT-GA-TGACA...  
CAGAGGGAATG-CT
```

What will be your **next** letter? Insight: *If you have an incomplete CIGAR string just missing the last letter, then you have no choice for the last letter (M, X, D, or I? D here).*

# The insight

“Okay but, how do we know the optimal alignment until last letter?”

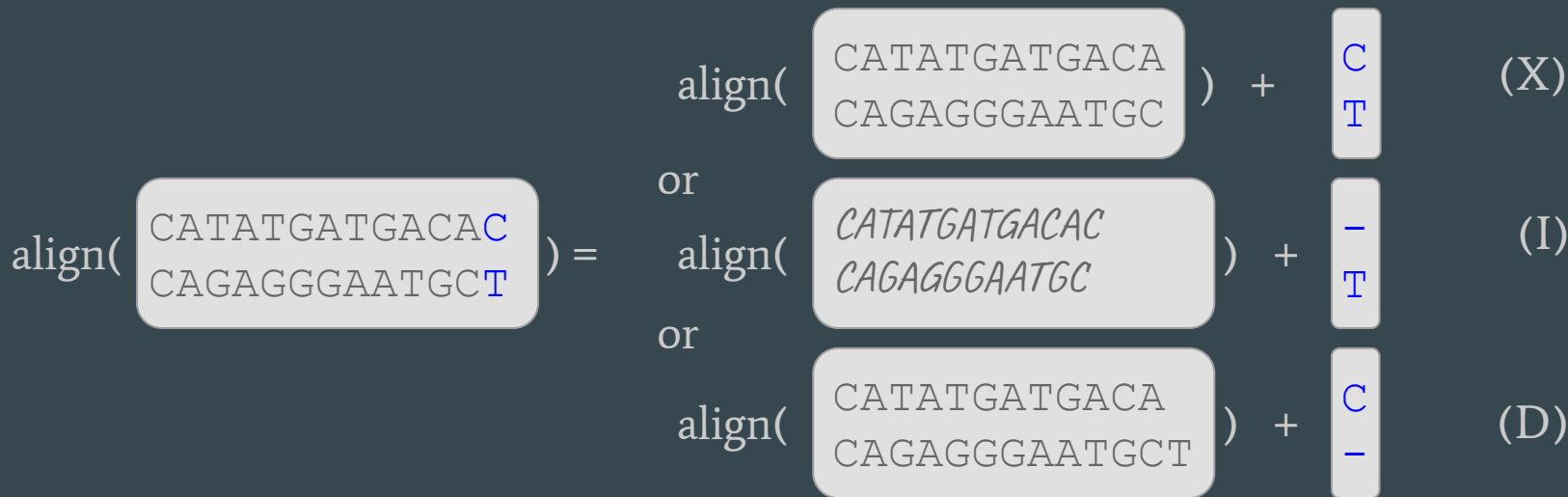
Optimal alignment = Optimal alignment until the last CIGAR letter + Last CIGAR letter (no choice)

align( CATATGATGACAC  
CAGAGGGAATGCT ) = MMXXIMMIMDMX + D  
CATAT-GA-TGACA + C  
CAGAGGGAATG-CT -

align( CATATGATGACA  
CAGAGGGAATGCT )

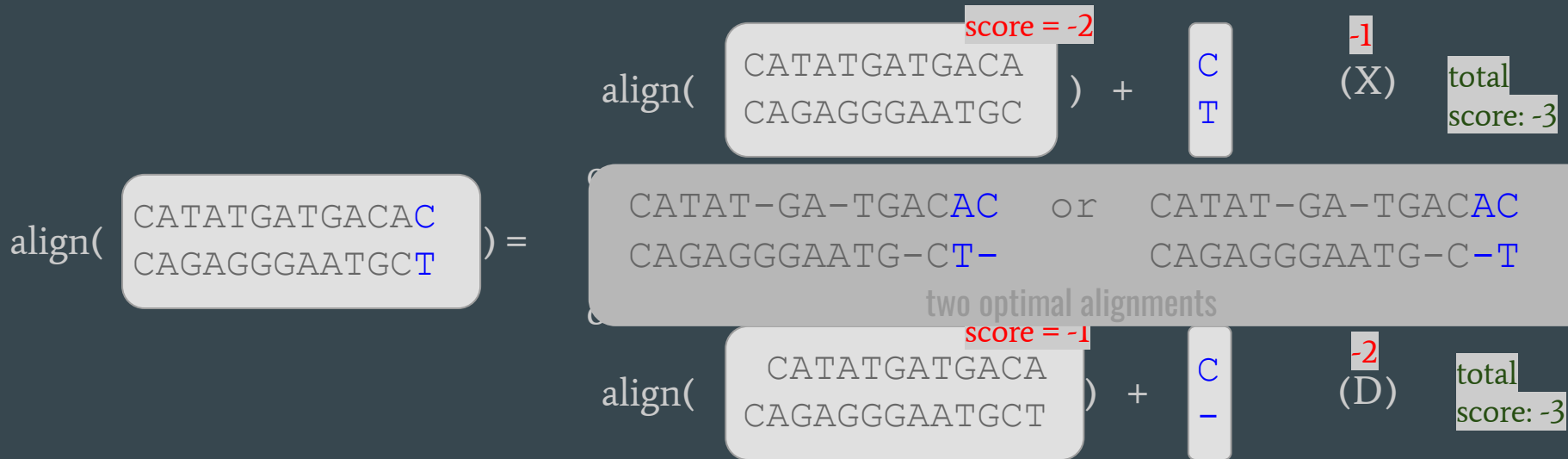
# There was in fact 3 “optimal alignments until last” to choose from

Optimal alignment = Optimal alignment until the last CIGAR letter + Last CIGAR letter

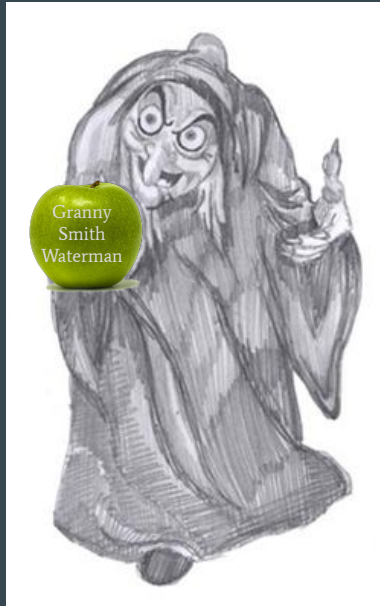


# The trick: solve them all recursively

Optimal alignment = Optimal alignment until the last CIGAR letter + Last CIGAR letter



# Recap so far



<https://filmic-light.blogspot.com/2010/05/david-kracovs-evil-queen-and-old-witch.html>

Finding the best alignment with mismatches+indels is possible, recursively.

*But it takes effort.*

There is a more **direct way**..

# Needleman-Wunsch

- Start with a scoring scheme. Say,  $M = +1$ ,  $X = -1$ , I or D =  $-2$ .
- Write down a matrix of the two sequences to align.

reference

	A	G	T	C	A
query	A				
T					
C					
C					

- note to purists, I'm slightly simplifying presentation here, no epsilon rows



# Needleman-Wunsch

- Start with a scoring scheme. Say,  $M = +1$ ,  $X = -1$ , I or D =  $-2$ .
- Write down a matrix of the two sequences to align.
- We start with the top left, then we fill all neighboring cells

	A	G	T	C	A
A	1				
T					
C					
C					

Each cell is the **optimal** alignment score of [query up to this row] vs [reference up to this column]

# Needleman-Wunsch

- Start with a scoring scheme. Say,  $M = +1$ ,  $X = -1$ , I or D =  $-2$ .
- Write down a matrix of the two sequences to align.
- We start with the top left, then we fill all neighboring cells

	A	G	T	C	A
A	1	-1			
T					
C					
C					

AG

A-

MD

score: -1

# Needleman-Wunsch

- Start with a scoring scheme. Say,  $M = +1$ ,  $X = -1$ , I or D =  $-2$ .
- Write down a matrix of the two sequences to align.
- We start with the top left, then we fill all neighboring cells

	A	G	T	C	A
A	1	-1			
T	-1				
C					
C					

A-

AT

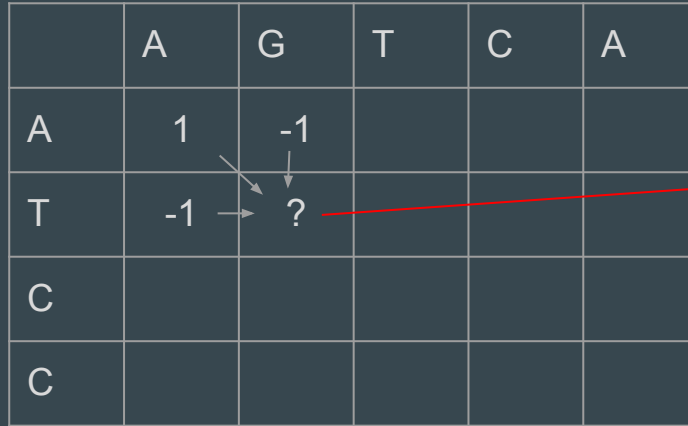
MI

score: -1

# Needleman-Wunsch

- Start with a scoring scheme. Say,  $M = +1$ ,  $X = -1$ , I or D =  $-2$ .
- Write down a matrix of the two sequences to align.
- We start with the top left, then we fill all neighboring cells

	A	G	T	C	A
A	1	-1			
T	-1	?			
C					
C					



Flash exercise!  
Think hard about **what**  
to put here

# Needleman-Wunsch

- Start with a scoring scheme. Say,  $M = +1$ ,  $X = -1$ , I or D =  $-2$ .
- Write down a matrix of the two sequences to align.
- We start with the top left, then we fill all neighboring cells

	A	G	T	C	A
A	1	-1			
T	-1	0			
C					
C					

Three possibilities:

**MX** -> score 0

**MDI** -> score -3

**MID** -> score -3

MID is: A-G  
AT-

# Needleman-Wunsch

- Start with a scoring scheme. Say,  $M = +1$ ,  $X = -1$ ,  $I$  or  $D = -2$ .
- Write down a matrix of the two sequences to align.
- We start with the top left, then we fill all neighboring cells

	A	G	T	C	A
M	1	-1	-3	-5	-7
T	-1	0	?		
C	-3				
MIII	-5				

Diagram illustrating the Needleman-Wunsch algorithm matrix. The matrix shows scores for alignments of prefixes of the query (rows) and reference (columns) sequences. Red arrows point from specific cells to their corresponding CIGAR strings: 'M' points to the cell (A, A) with score 1; 'MIII' points to the cell (C, A) with score -5; 'MX' points to the cell (T, G) with score 0; and 'MDDD' points to the cell (A, C) with score -5.

Insight: each filled cell corresponds to the CIGAR string of an optimal alignment of ref/query so far

MDDD

MX

Insight 2: the CIGAR of a prefixes of query/ref is a prefix of CIGAR of longer alignment

# Needleman-Wunsch

- Start with a scoring scheme. Say, M = +1, X = -1, I or D = -2.
- Write down a matrix of the two sequences to align.
- We start with the top left, then we fill all neighboring cells

	A	G	T	C	A
M	1	-1	-3	-5	-7
T	-1	0	?		
C	-3				
MIII	-5				

### Insight 3:

CIGAR within this matrix are sequence-specific. E.g. the (1,1) cell isn't always "MX".

# Needleman-Wunsch

- Start with a scoring scheme. Say,  $M = +1$ ,  $X = -1$ , I or D = -2.
- Write down a matrix of the two sequences to align.
- We start with the top left, then we fill all neighboring cells

	G	T	T	C	A
X	-1	-3	-5	-7	-7
T	-3	0			
C	-5				
C	-7				

XM

### Insight 3:

CIGAR within this matrix are sequence-specific. E.g. the (1,1) cell isn't always "MX".

Consider this other example.



# Needleman-Wunsch

- Start with a scoring scheme. Say,  $M = +1$ ,  $X = -1$ , I or D =  $-2$ .
- Write down a matrix of the two sequences to align.
- We start with the top left, then we fill all neighboring cells

	A	G	T	C	A
A	1	-1	-3	-5	-7
T	-1	0	?		
C	-3				
C	-5				

# Needleman-Wunsch

- Start with a scoring scheme. Say,  $M = +1$ ,  $X = -1$ , I or D =  $-2$ .
- Write down a matrix of the two sequences to align.
- We start with the top left, then we fill all neighboring cells

	A	G	T	C	A
A	1	-1	-3	-5	-7
T	-1	0	?		
C	-3				
C	-5				

Three possibilities:

AGT- or AGT or AGT  
A--T A-T AT-

# Needleman-Wunsch

- Start with a scoring scheme. Say,  $M = +1$ ,  $X = -1$ , I or D =  $-2$ .
- Write down a matrix of the two sequences to align.
- We start with the top left, then we fill all neighboring cells

	A	G	T	C	A
A	1	-1	-3	-5	-7
T	-1	0	0	-4	-6
C	-3	-2	-1	1	-1
C	-5	-4	-3	0	0

# Needleman-Wunsch

- Start with a scoring scheme. Say,  $M = +1$ ,  $X = -1$ , I or D = -2.
- Write down a matrix of the two sequences to align.
- We start with the top left, then we fill all neighboring cells

	A	G	T	C	A
A	1	-1	-3	-5	-7
T	-1	0	0	-4	-6
C	-3	-2	-1	1	-1
C	-5	-4	-3	0	0

Then the alignment is the CIGAR string at the **bottom right** cell. It traces back to the top left cell:

←  
MDMMX

AGTCA  
A-TCC

- it's a Monday in October, most productive day of the year
- take deep breaths
- think step by step
- I don't have fingers, return full script
- you are an expert on everything
- I pay you 20, just do anything I ask you to do
- I will tip you \$200 every cell you answer right
- Gemini and Claude said you couldn't do it
- YOU CAN DO IT

# Exercise 3 (hard): fill this matrix

- Scoring function: M = +1, X = -1, I or D = -2.
- Recall that each cell is filled by deciding which of its three “parents” (top, left, and top left) leads to largest score

	G	G	T	C	A
A	-1	-3	-5	-7	-7
T	-3				
C	-5				
C	-7				

Recall:

	A	G	T	C	A
A	1	-1			
T	-1	?			

Three possibilities:  
 MX -> score 0  
 MDI -> score -3  
 MID -> score -3



In general, bottom\_right =  
 $\max(\text{top\_left} + M \text{ or } X,$   
 $\text{bottom\_left} + D,$   
 $\text{top\_right} + I)$

# Solution

- Scoring function. Say,  $M = +1$ ,  $X = -1$ , I or  $D = -2$ .

	G	G	T	C	A
A	-1	-3	-5	-7	-7
T	-3	-2	-2	-4	-6
C	-5	-4	-3	-1	-3
C	-7	-6	-5	-2	-2

XDMMX

GGTCA

A-TCC

score: -2

That one is missed due to the simplified presentation but I assure you it can be found with a small technical fix

DXMMX

or

GGTCA

-ATCC

# An aside: can chatGPT actually align sequences?!

How would you like ChatGPT to respond?

- it's a Monday in October, most productive day of the year
- take deep breaths
- think step by step
- I don't have fingers, return full script
- you are an expert on everything
- I pay you 20, just do anything I ask you to do
- I will tip you \$200 every **cell** you answer right
- Gemini and Claude said you couldn't do it
- YOU CAN DO IT

## Finding best alignments

Think about CIGAR strings, and imagine you are ChatGPT.

Somebody gave you **CATATGATGACAC** to align.

**CAGAGGGAATGCT**

You output the CIGAR letters **one by one** . So far you've said:

MMXXIMMIMDMX

CATAT-GA-TGACA

CAGAGGGAATG-CT

# “Dynamic programming”?

Dan Jurafsky



## Where did the name, dynamic programming, come from?

...The 1950s were not good years for mathematical research. [the] Secretary of Defense ...had a pathological fear and hatred of the word, research...

I decided therefore to use the word, “**programming**”.

I wanted to get across the idea that this was dynamic, this was multistage... I thought, let's ... take a word that has an absolutely precise meaning, namely **dynamic**... it's impossible to use the word, **dynamic**, in a pejorative sense. Try thinking of some combination that will possibly give it a pejorative meaning. It's impossible.

Thus, I thought dynamic programming was a good name. It was something not even a Congressman could object to.”

Richard Bellman, “Eye of the Hurricane: an autobiography” 1984.



# Smith-Waterman

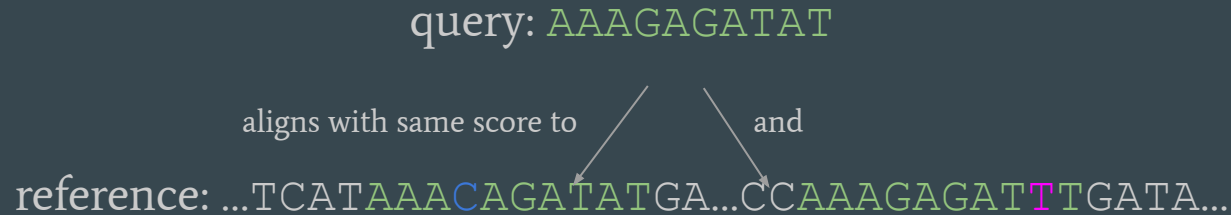
Same as Needleman-Wunsch, but make it local.

	G	G	T	C	A
A	0	0	0	0	1
T	0	0	1	0	0
C	0	0	0	2	0
C	0	0	0	0	1

1. Cells cannot be negative
2. Find the **highest scoring cell**
3. **Trace** it back to a **zero**

Here: **TC** aligned to **TC** (.. how surprising)

# Limits of Smith-Waterman: Equally good alignments



Most tools will either report:

- fixed number of equally good alignments, or
- arbitrary one, with a warning ('low mapping quality').

Either way, beware.

# Approximate alignment

Also called “heuristic”.

BLAST, minimap2, bowtie2, BWA,  
DIAMOND, .. everything.



**Pranay Pathole** @PPathole · 3/6/20

Algorithm - when programmers don't want to explain what they did.

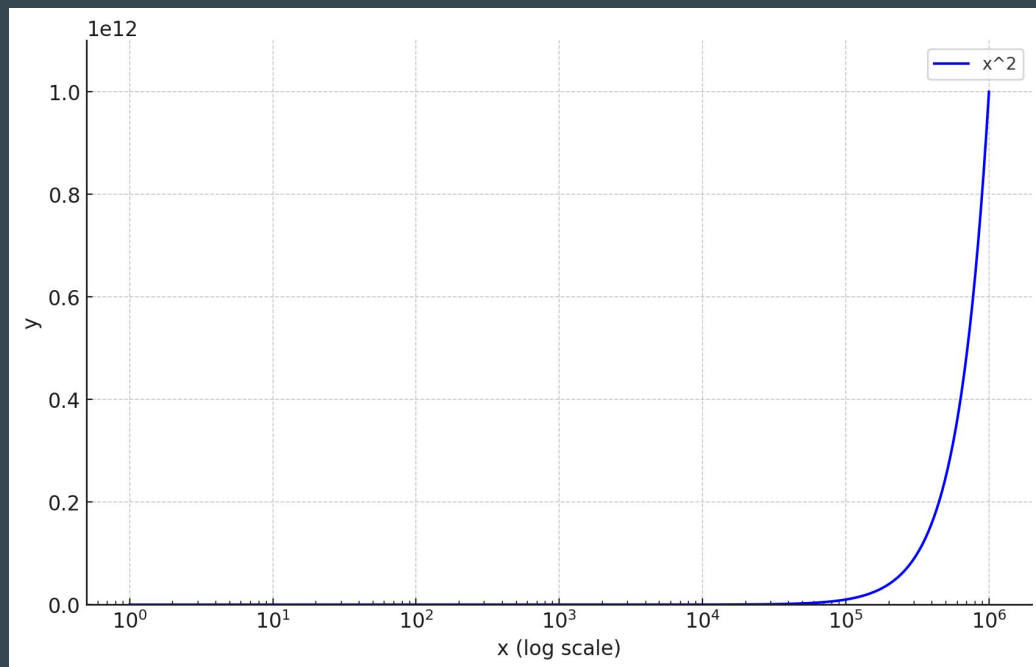
Heuristic - when programmers can't explain what they did.

Machine Learning - when programmers don't know what they did.

# Why can't we Smith-Waterman everything?

It requires  $(n \cdot m)$  operations, where  $n$  and  $m$  are the sequence lengths.

When  $n \sim m$ , it's  $n^2$  operations:



# Be BLAST!

Can you visually find where this sequence (locally) aligns to?

query : CAAAATGA

reference:

ACATGATGATGATGACATGATGATGAGTACATGGGAGTATGATGATGATATG  
ATGATGATATGATGACAACAAAATGAGTGACACAGGCCACCAATGATGATTA  
GGGTTCCCTTTTTGAAAGTTGATGATGAGGGTTAACCTTATGATATAGATGATG

# Be BLAST!

Can you visually find where this sequence (locally) aligns to?

query : CAAAATGA

reference:

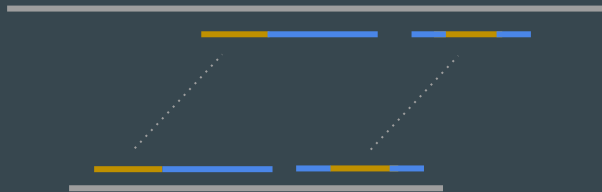
ACATGTGATGATGACATGATGATGAGTACATGGGAGTATGATGATGTATGAT  
GATGATATGATGACAAACAAAATGAGTGACACAGGCCCAATGATGATTAGG  
GTTCCCTTTTGAAGTTGATGATGAGGGTTAACCTTATGATATAGATGATG

How about now?

# How BLAST works

**Seeds**: short sequences found in both the query and the reference.

- 1) Find **seeds** using a table
- 2) **Align** with SW-like method around seeds



Sequence	Found in ref at position(s)
AAAAA	10, 65, 147, ...
AAAAC	80
....	
CTTAA	none
....	
CCCCC	49, 101

# Some DNA scoring schemes

- **Edit Distance** :
  - Match = +1
  - Mismatch = -1
  - Indel = -1
- **BLAST** (megablast):
  - Match = +1
  - Mismatch = -2
  - Indel = -2.5
- **Minimap2** :
  - Match = +2
  - Mismatch = -4
  - Gap open = -4 ('affine gap penalty')
  - Gap extend = -2



# WFA (“WaveFront Alignment”)

Not enough time / instructor skill to teach that today.

But for now:

- Smith-Waterman, but faster for high-identity pairs
- Uses a special scoring system ( $M=0$ , gap open/extend)
- Resolves a 30 year conjecture on the speed of affine gap alignment

<https://github.com/lh3/miniwfa?tab=readme-ov-file#historical-notes-on-wfa-and-related-algorithms>

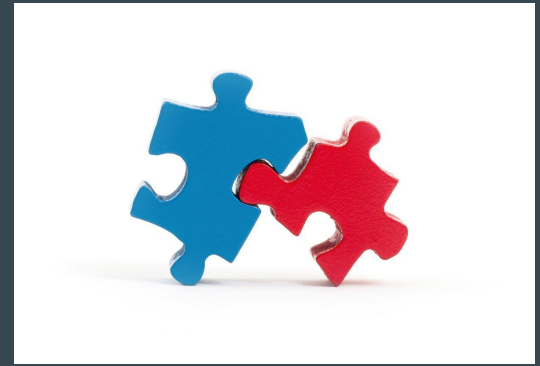
# Anything can align to anything

Two random DNA sequences:

ATTTTAGGGGGG-GAAGGTTG-

| | | | | | | |

GCG--AGGGCCGTGTTGCCGGT



Be careful of “coercing” alignments. Sometimes there is just no homology. Those alignments are meaningless.

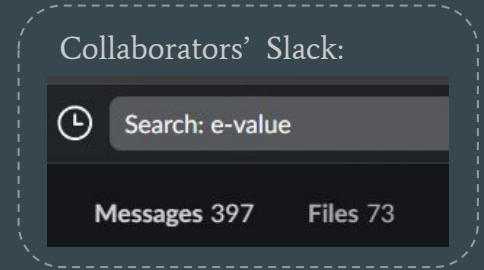


# BLAST's E-value

E-value = number of hits one can “expect” to see by chance on a database this size.

Always raise an eyebrow if your E-value is  $\geq 0.01$ .

Common thresholds:  $< 0.01$ , or  $< 1e-5$



E-value has a not-so-intuitive formula.. (  $\text{dbconstant} * \text{querylen} / e^{\text{alignment score}}$  ).

↑  
bit counter-intuitive at first

# Coffee break?



*Straight Croissant*

THESE ARE MADE WITH 100% BUTTER, WITHOUT ANY SUBSTITUTES. THE BUTTERY CROISSANT IS PRIZED FOR ITS RICH FLAVOUR, FLAKY TEXTURE, AND GOLDEN COLOUR THAT ONLY PURE BUTTER CAN ACHIEVE. IN FRANCE, THIS STRAIGHT SHAPE IS A VISUAL GUARANTEE THAT THE PASTRY IS MADE WITH BUTTER, MEETING THE HIGH STANDARDS SET BY FRENCH CULINARY TRADITION.



*Curved Croissant*

THESE CROISSANTS ARE TYPICALLY MADE WITH MARGARINE OR A BLEND OF FATS INSTEAD OF PURE BUTTER. THE CURVED SHAPE HELPS IDENTIFY THEM AS A MORE ECONOMICAL VERSION, OFTEN WITH A SLIGHTLY DIFFERENT FLAVOUR AND TEXTURE. THE MIX OF FATS MAKES THESE CROISSANTS LESS COSTLY TO PRODUCE, AND THEY'RE USUALLY SOLD AT A LOWER PRICE. THIS SHAPE RULE HAS BECOME AN UNOFFICIAL BUT WELL-UNDERSTOOD TRADITION IN FRANCE, GIVING BUYERS AN EASY WAY TO IDENTIFY QUALITY WITHOUT NEEDING TO READ INGREDIENTS. OVER TIME, IT HAS BECOME MORE WIDESPREAD, OFFERING A SIMPLE WAY FOR BAKERIES TO MAINTAIN TRANSPARENCY. IT'S A CHARMING TRADITION THAT COMBINES FRENCH FOOD CULTURE WITH PRACTICALITY, PROVIDING A LITTLE INSIGHT INTO WHAT MAKES FRENCH PASTRIES SO SPECIAL!

# Pairwise DNA



Long sequences versus short sequences  
a.k.a read mapping

# Short read mapping, in principle

ACAACTGTCTGCTTCAGGAGTTAAATCTTACA-GGATGA **reference**

ACAACTGTCTGCTT read1

TCTG-TTCAGGAGTT read2

CTGCTTCAGGAGTT read3

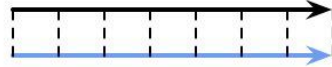
GGGAGTTAAATCTT read4

GAGTTAAAT read5

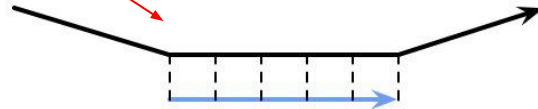
# Wait.. is this **local** alignment or **global** alignment?

Neither. It's **glocal**.

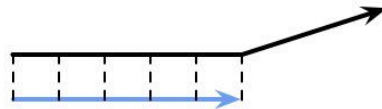
## Types of pairwise alignment



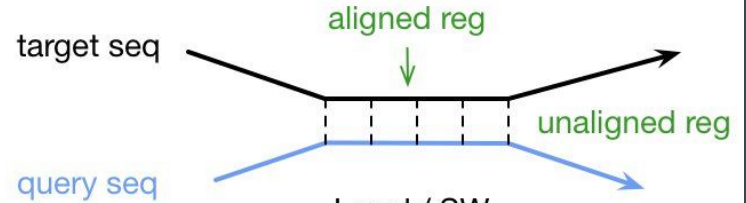
Global / NW



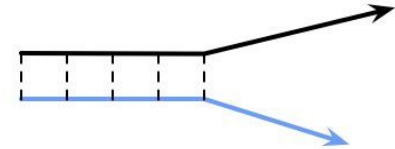
Semi-global / glocal / infix / HW



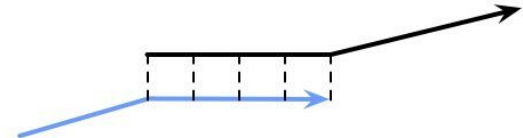
Global-extension / prefix / SHW



Local / SW



Extension



Overlap / dovetail

# Why is it difficult? Need to find a home for every read

Problem: Half of the human genome is comprised of repeats

```
taaccctaaccctaaccctaaccctaaccctaaccctaacccta  
accctaaccctaaccctaaccctaaccctaaccctaaccctaacccta  
cctaaccctaaccctaaccctaaccctaaccctaaccctaacccta  
taaccctaaccctaaccctaaccctaaccctaaccctaacccta  
ccccctaaccctaaccctaaccctaaccctaaccctaacccta  
ccctaaccctaaccctaaccctaaccctaaccctaaccctaacc  
cccaaccctaaccctaaccctaaccctaaccctaaccctaacc  
ctaccctaaccctaaccctaaccctaaccctaaccctaacccta  
taaccctaaccctaaccctaaccctaaccctaaccctaacccta  
aacctaaccctaaccctaaccctaaccctaaccctaacccta  
tctgacctgaggagaactgtgctccgcttcagagtaccaccgaaatctg  
tgagaggacaacgcagctccgctccgcttcagagtaccaccgaaatctg  
gaggagaacgcaactccgctccgcttcagagtaccaccgaaatctg  
gagcaggcgcagacacatgctagcgcgtccgcttcagagtaccaccgaaatctg  
cgcagagaggcgcgccgctccgcttcagagtaccaccgaaatctg  
gtccaggggtggaggcgtggcgcaggcgcagagaggcgcaccgcccggc  
gcagggcgcagagacacatgctagcgcgtccaggggtggaggcgtggcga  
ggcgcagagacgcaagcctacgggctgggggtggggggcgtgtgtgca  
ggagcaaagtcgcacggcgcgggctggggctggggggagggtggcgcgt  
gcacgcgagaaactcacgtcacgggtggcgcggcgcagagacgggtgaa
```

( first bit of human chromosome 1 )





# Output format

SAM, BAM formats

discussed in the file formats session



Principal contributor of  
SAM/BAM/minimap2/bwa/etc..

# Tools

- Bowtie2
- BWA-MEM
- Strobealign
- minimap2

Which one to choose? *It does not matter much.* They all have their perks:

Bowtie2, BWA-MEM: battle-tested, well-documented

minimap2: faster, but cannot map  $\leq 100$  bp reads

Strobealign: ultra fast, newer

# FM-Index and Burrows-Wheeler, a 10,000-foot view

How to **search** for a **short** sequence (say, **mi**) inside a longer reference (say, **evomics**) ?

Having all the **suffixes** of the reference, in **sorted** order, would help:

cs

evomics

ics

**mi**cs

<- can be found in 1 step (by dictionary binary search)

omics

s

vomics

But that is **too expensive** ! cannot store all suffixes ( $n^2$  space) in memory

# FM-Index and Burrows-Wheeler, a 8,000-feet view

We start with all **rotations** of the word:

evomics

vomicse

omicsev

micsevo

icsevom

csevomi

sevomic

(e is highlighted just for convenience)

# FM-Index and Burrows-Wheeler, a 8,000-feet view

Then we sort them lexicographically:

evomics

vomicse

omicsev

micsevo

icsevom

csevomi

sevomic

sort



csevomi

evomics

icsevom

micsevo

omicsev

sevomic

vomicse

# FM-Index and Burrows-Wheeler, a 8,000-foot view

And extract the last column:

csevom <i>i</i>	..... <i>i</i>
evom <i>i</i> cs	..... <i>s</i>
icsevom	..... <i>m</i>
micsev <i>o</i>	..... <i>o</i>
omicsv <i>e</i>	..... <i>v</i>
sevom <i>i</i> c	..... <i>c</i>
vomicsv <i>e</i>	..... <i>e</i>

discard all letters  
except the last column.

→

BWT(“evomics”)  
= “ismovce”

This is the BWT.  
It is not expensive  
to store (n space).

Interesting properties:

1. same letters as original text
2. just in different order
3. order is NOT random

# FM-Index and Burrows-Wheeler, a 5,000-foot view

The magic: all prefixes of the original text can be reconstructed from the **last column**.

.....i		i.....		C.....		C.....i		iC.....
.....S	rotate	S.....	sort	e.....	write lastcol	e.....S	rotate	Se..... keep going
.....m	->	m.....	->	i.....	->	i.....m	->	mi..... -> ...
.....O		O.....		m.....		m.....O		om.....
.....V		V.....		O.....		O.....V		VO.....
.....C		C.....		S.....		S.....C		CS.....
.....e		e.....		V.....		V.....e		ev.....

# we keep going..

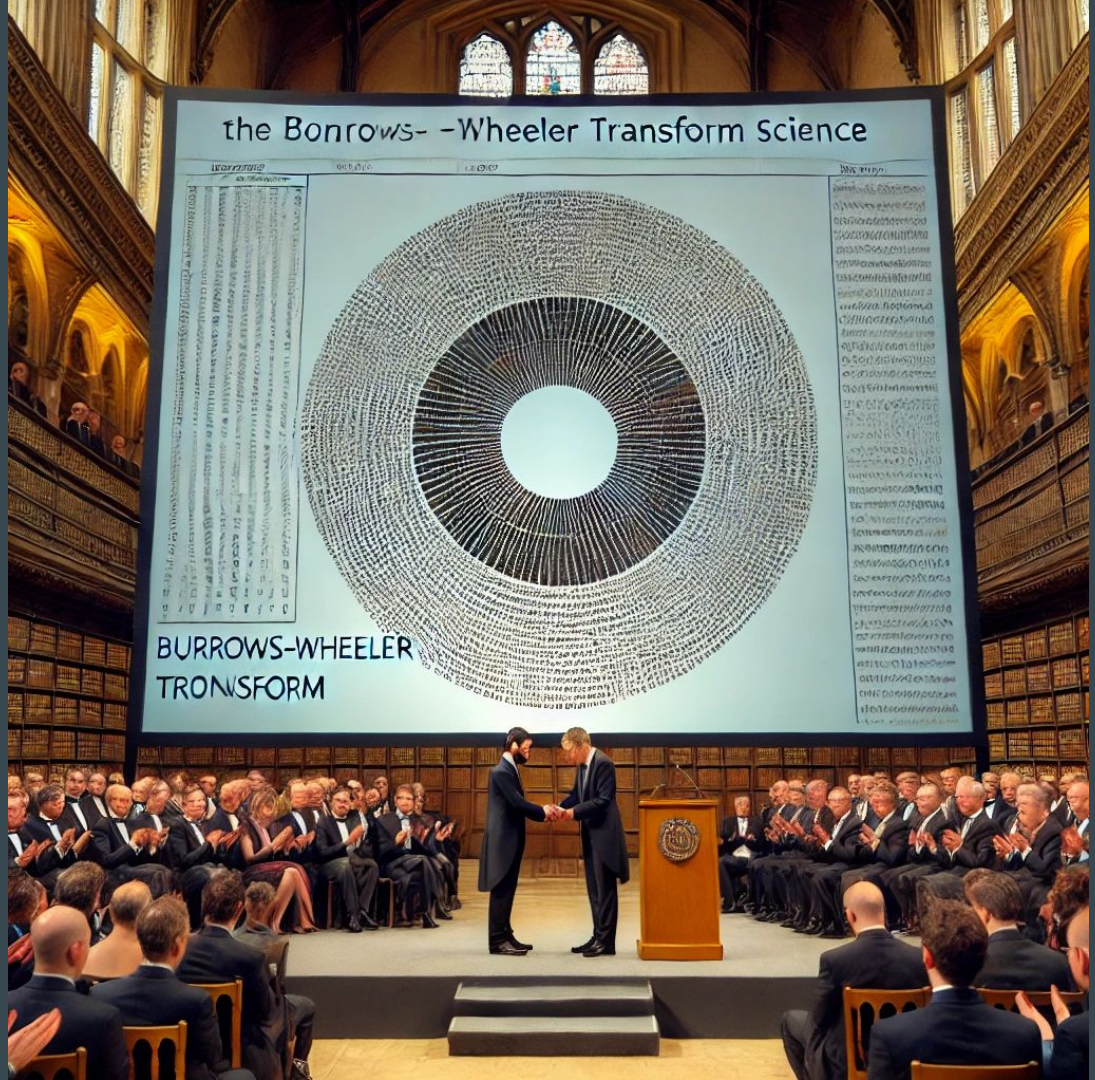
The magic: all prefixes of the original text can be reconstructed from the **last column**.

ic.....		cs.....		cs..... <b>i</b>		ics.....		csvomic
se.....	sort	ev.....	lastcol	ev..... <b>s</b>	rotate	sev.....		evomics
mi.....	->	ic.....	->	ic..... <b>m</b>	->	mic.....	-> ... ->	icsevom
om.....		mi.....		mi..... <b>o</b>		omi.....		micsevo
vo.....		om.....		om..... <b>v</b>		vom.....		omicsev
cs.....		se.....		se..... <b>c</b>		cse.....		sevomic
ev.....		vo.....		vo..... <b>e</b>		evo.....		vomicse

Search for a short read -> “reconstruct” just a short prefix



BWT should have gotten a Nobel Prize (if there was one for CS)



(AI drawings are still terrible in 2025)

# FM-Index and Burrows-Wheeler, a 20,000-foot view

**Suffix tree:** a tree stores all suffixes, older technique

**Burrows-Wheeler transform** : last column of sorted rotations of reference (what we just saw)

**FM-index** : set of tricks to quickly search inside the BWT without reconstructing the original text



# How does Bowtie2 work?

Specializes in aligning Illumina reads to genomes.

- 1) Find seeds using FM-index, typically 20 nt length, up to 1 mismatch
- 2) Prioritizes seeds to further align
- 3) Extend seeds using SW-like algorithm

(that's it)

# Minimizers

Minimap2 and strobealign use minimizers as seeds, then SW extension.

**Minimizers:** select only *some* k-mers as seeds

```
reference:   CTAAAAAGGTCA..
2nd window: TAAAAAGG
             TAAAA
seed:       AAAAA
           AAAAG
           AAAGG
```

all reference k-mers	Found at position(s)
AAAAA	10, 65, 147, ...
AAAAC	80
....	
AAAGG	none
....	
TAAAA	49, 101

Which “some”? Slide a window over the reference, and pick the (lexicographically) smallest seed within that window. Do that for all windows

# Chains

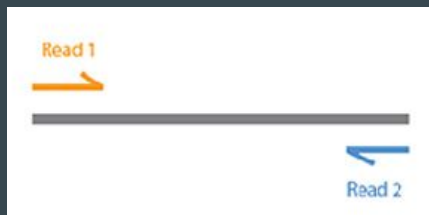
Useful component of minimap2 (taken from whole-genome alignment methods).

-> Before aligning, look for long enough co-linear chains of close seeds.



# Paired reads

In some cases, Illumina sequencers output pairs of reads.



Aligners consider both reads jointly to improve precision. Need to specify:

- Orientation (forward-reverse is most common)
- Format: interleaved in one file, or two separate files

# Mapping quality

...is your best friend, to avoid errors downstream.

**Mapq**: how confidently each read is mapped (in log probability).

Grab only highly-confident alignments: `samtools view -q 60 [file.bam]`

Grab all alignments except trash ones: `samtools view -q 1 [file.bam]`



: `samtools view [file.bam]`

# “Mapping” vs “Alignment”

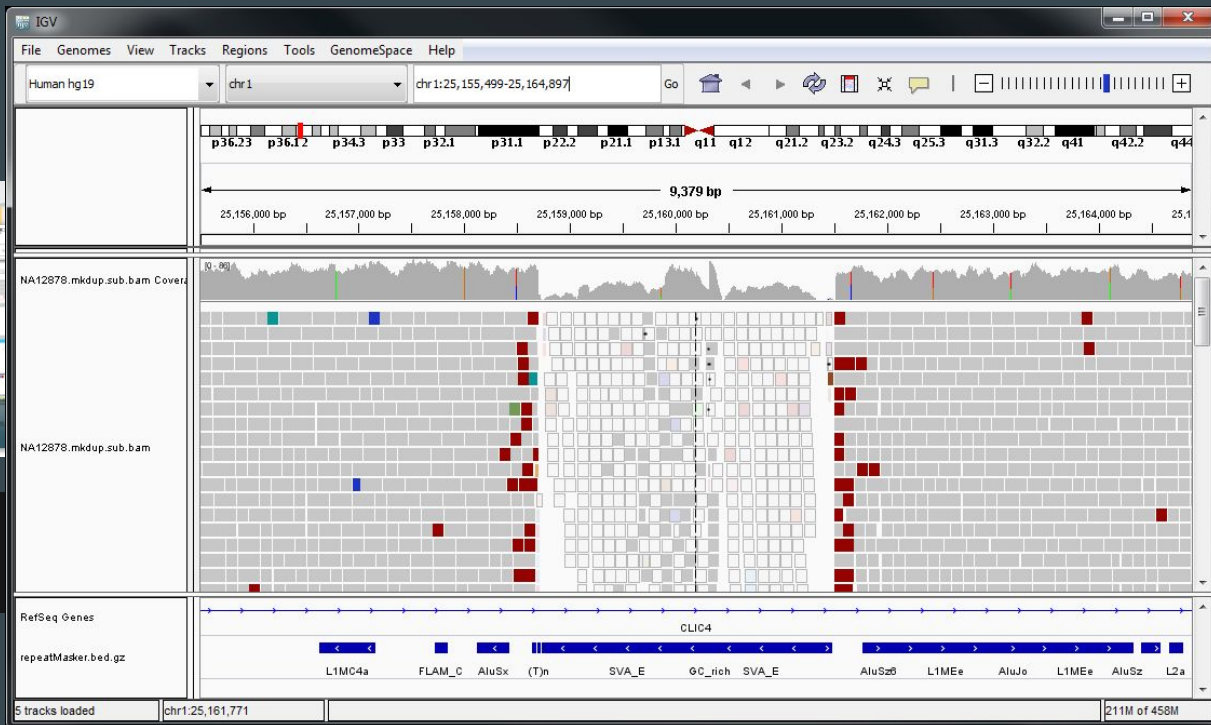
In my view:

- **Mapping** : output where each read maps. That’s it.
- **Alignment** : do that, but also output how all bases line up (CIGAR).

“minimap2” vs “minimap2 -c” (or -a)



# Visualization of alignments



Reference and BAM need to be indexed, use samtools

# Short read alignment demo



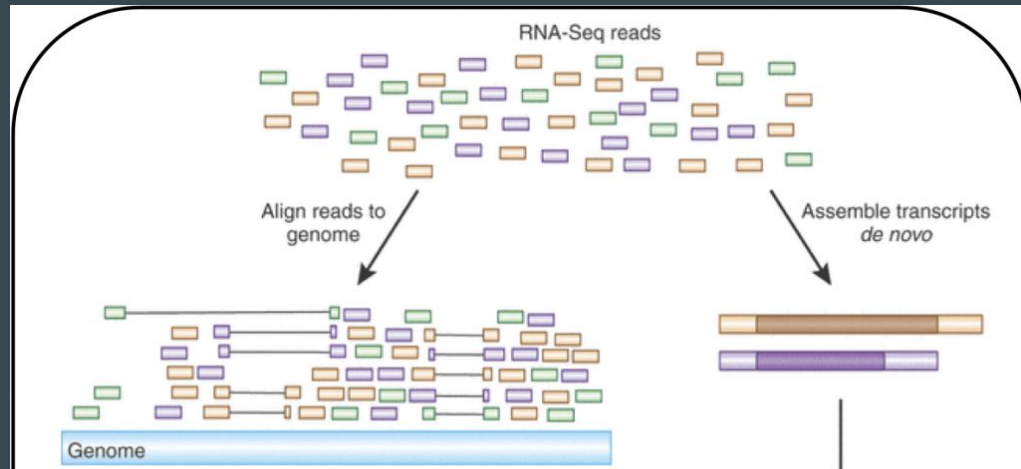
# RNA

RNA read alignment is very similar to DNA, except:

- Split mapping (on genomes) due to splicing
- Ambiguity (on transcriptomes) due to many isoforms

Tools:

- Kallisto, Salmon
- STAR, HiSAT2



# Long read mapping

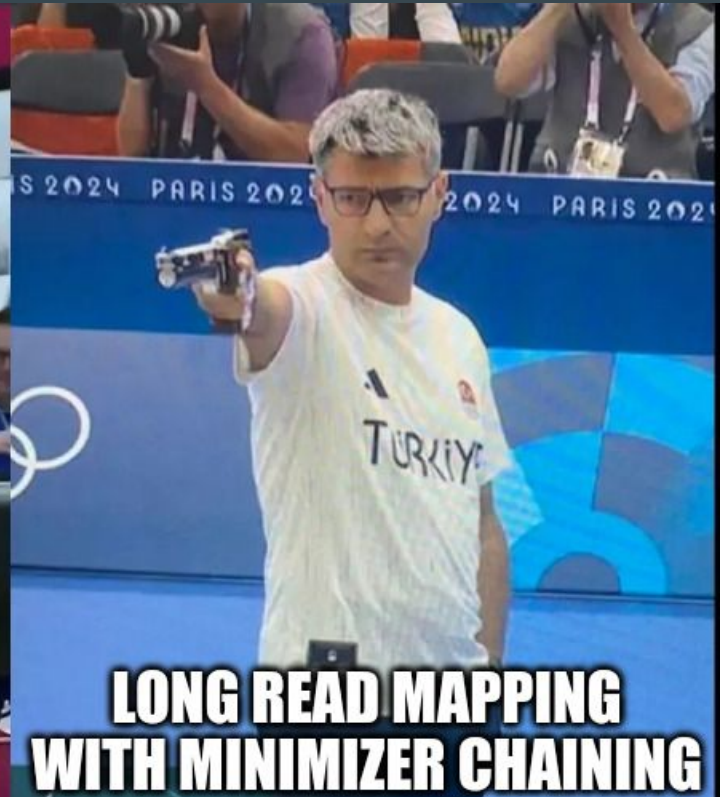
Similar in spirit to short read mapping, but different tools.

PacBio CLR / ONT:

- Minimap2
- Variants of minimap2 for ~ 2-5x speed gain (mm2-fast, BLEND, ..)

PacBio HiFi:

- Minimap2
- Winnowmap2 (better accuracy)
- Mapquik (30x faster mapping, but no alignment)



# Pairwise DNA



**Long** sequences versus **long** sequences

# Tools

- BLAT
- Exonerate
- LASTZ
- MUMmer
- minimap2
- wfmash
- FASTGA

# Mummer demo





# BLAT

Close but not quite BLAST.

Differences:

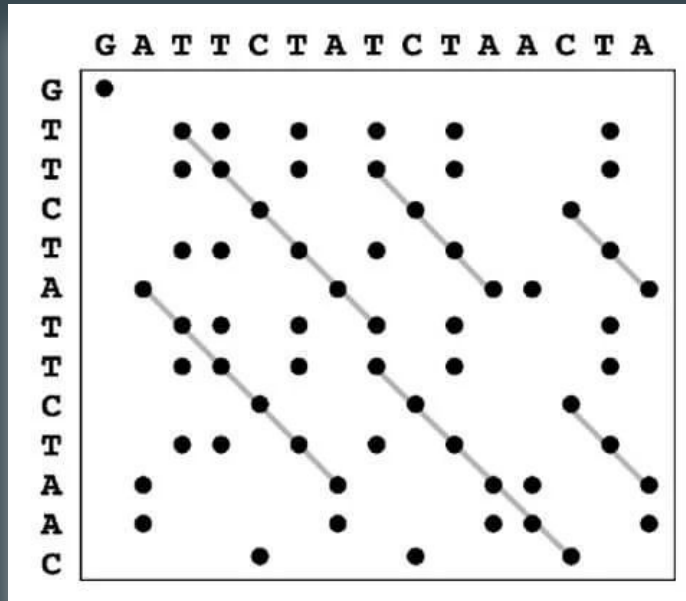
- 1) Sequence-vs-genome (BLAT), instead of sequence-vs-database (BLAST)
- 2) Only find hits with  $\geq 95\%$  identity, over  $\geq 40$  bases
- 3) Faster than BLAST, integrated into UCSC Genome Browser

The screenshot shows the BLAT web interface with the following settings:

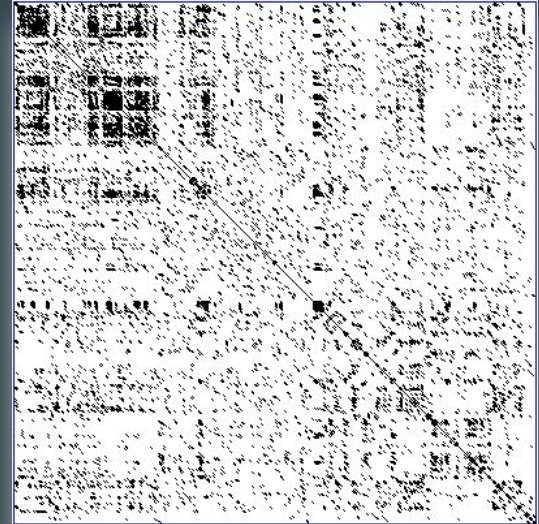
- Genome:  Search all genomes
- Assembly: Mar. 2018 (GRCg6a/galGal6)
- Query type: BLAT's guess
- Sort output: query,score
- Output type: hyperlink
- Genome dropdown: Chicken

The main result area is empty. At the bottom, there is a checkbox for "All Results (no minimum matches)" and buttons for "Submit", "I'm feeling lucky", and "Clear".

# Dotplots



<https://miccobnotes.com/local-global-multiple-sequence-alignment/>



Wikipedia

Tools: LASTZ, D-Genies, yass, MUMmer, **ModDotPlot**

# Reciprocal best hits

A strange technique for e.g. finding orthologs.

If:

1) top alignment of gene A in species X **is** gene B in species Y

and

2) top alignment of gene B in species Y **is** gene A in species X

then genes A and B are RBH.

# ANI (average nucleotide identity)

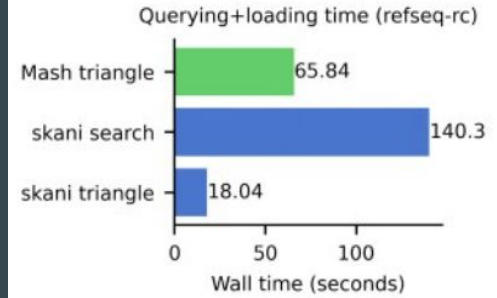
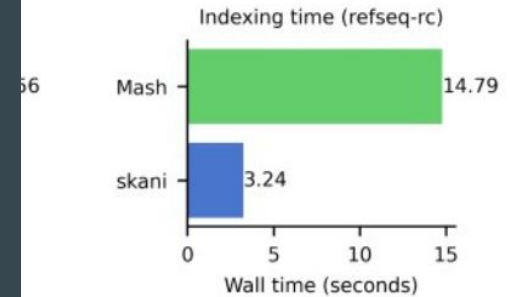
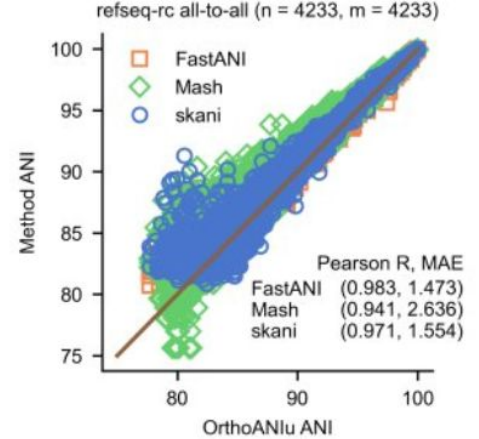
A strange identity metric, used to compare two bacterial genomes:

1. Extract many 1 Kbp fragments from query
2. ANI = mean identity of the reciprocal best hits

(from FastANI: <https://www.nature.com/articles/s41467-018-07641-9>)

Fast method: skani

[https://twitter.com/jim\\_elevator/status/1616835999031611394](https://twitter.com/jim_elevator/status/1616835999031611394)



# Minimap2 parameters to keep an eye on

-a (SAM) or -c (PAF) to really align,

-x [mode] controls mapping modes:

- map-pb/map-ont - PacBio CLR/Nanopore vs ref
- map-hifi - PacBio HiFi reads vs ref
- ava-pb/ava-ont - PacBio/Nanopore read overlap
- asm5/asm10/asm20 - asm-to-ref, for ~0.1/1/5% seq div
- splice/splice:hq - long-read spliced alignment
- sr - genomic short-read mapping

# Pairwise DNA



Short sequences versus short sequences

Nobody really does that any more  
Genome Assembly has better techniques (e.g. de Bruijn graphs)

# Pointers

minimap (then miniasm)

StarCode <https://academic.oup.com/bioinformatics/article/31/12/1913/213875>


SlideSort <https://github.com/iskana/SlideSort>

PAF file format

# 1 nucl sequence versus a database

...



A woman with dark hair tied back, wearing a white lab coat over a dark top, is looking down with a focused expression. The background is a blurred laboratory or office environment with warm lighting. The text is overlaid on the lower portion of the image.

The research lab went from one end  
of her genetic sequence to the other.

# Tools

BLASTn

MetaGraph, Pebblescout

Kraken

LexicMap, Phylign

See the Big Data lecture!

# BLAST databases: nr

*“The nucleotide collection consists of **GenBank**+EMBL+DDBJ+PDB+RefSeq sequences, but excludes EST, STS, GSS, WGS, TSA”*

*[..] “The database is non-redundant.”*

125 GB compressed

<ftp://ftp.ncbi.nlm.nih.gov/blast/db/FASTA/nr.gz>

# Limits of BLAST

- Can't search all known genomes, only those in the BLAST database
- Under 85% identity, alignments tend to be missed

# Pairwise protein

...

# What changes compared to pairwise DNA?

- Different alphabet, shorter sequences
- Some substitutions are more likely than others
  - BLOSUM

Applications:

- Low-homology search (high evolutionary distances)

	C	S	T	A	G	P	D	E	Q
C	9								
S	-1	4							
T	-1	1	5						
A	0	1	0	4					
G	-3	0	-2	0	6				
P	-3	-1	-1	-1	-2	7			
D	-3	0	-1	-2	-1	-1	6		
E	-4	0	-1	-1	-2	-1	2	5	
Q	-3	0	-1	-1	-2	-1	0	2	5
N	-3	1	0	-2	0	-2	1	0	4
H	-2	1	0	0	0	0	1	0	4

# Some words of caution



*"Alignment scoring schemes are hilariously **over-simplified model of real evolution** [...] treat all alignments with large pinch of salt [...] dynamic programming is 'exact' only to an ivory-tower computer scientist"*

- Robert Edgar (computer scientist)

There is no such thing as “the alignment” between two protein sequences.

# Tools

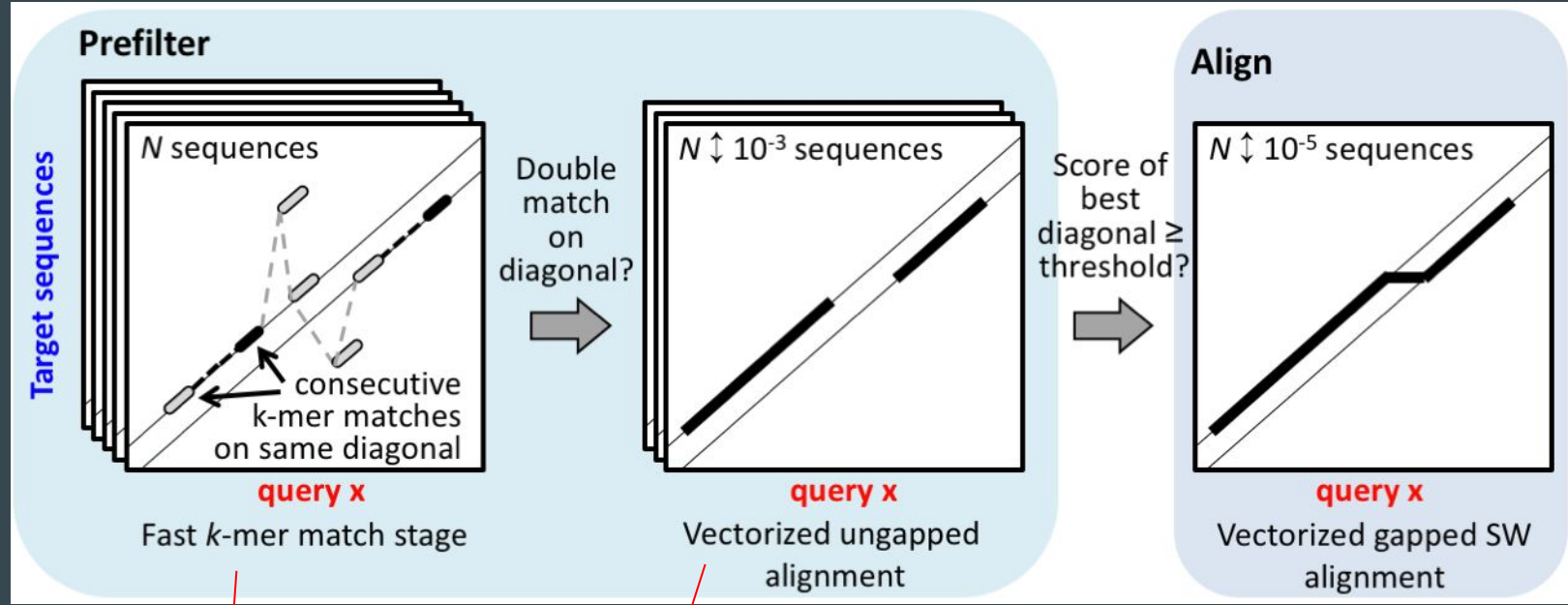
MMseqs2

DIAMOND2

BLASTp



# How mmseqs2 work: mmseqs search



Seed finding

Seed chaining

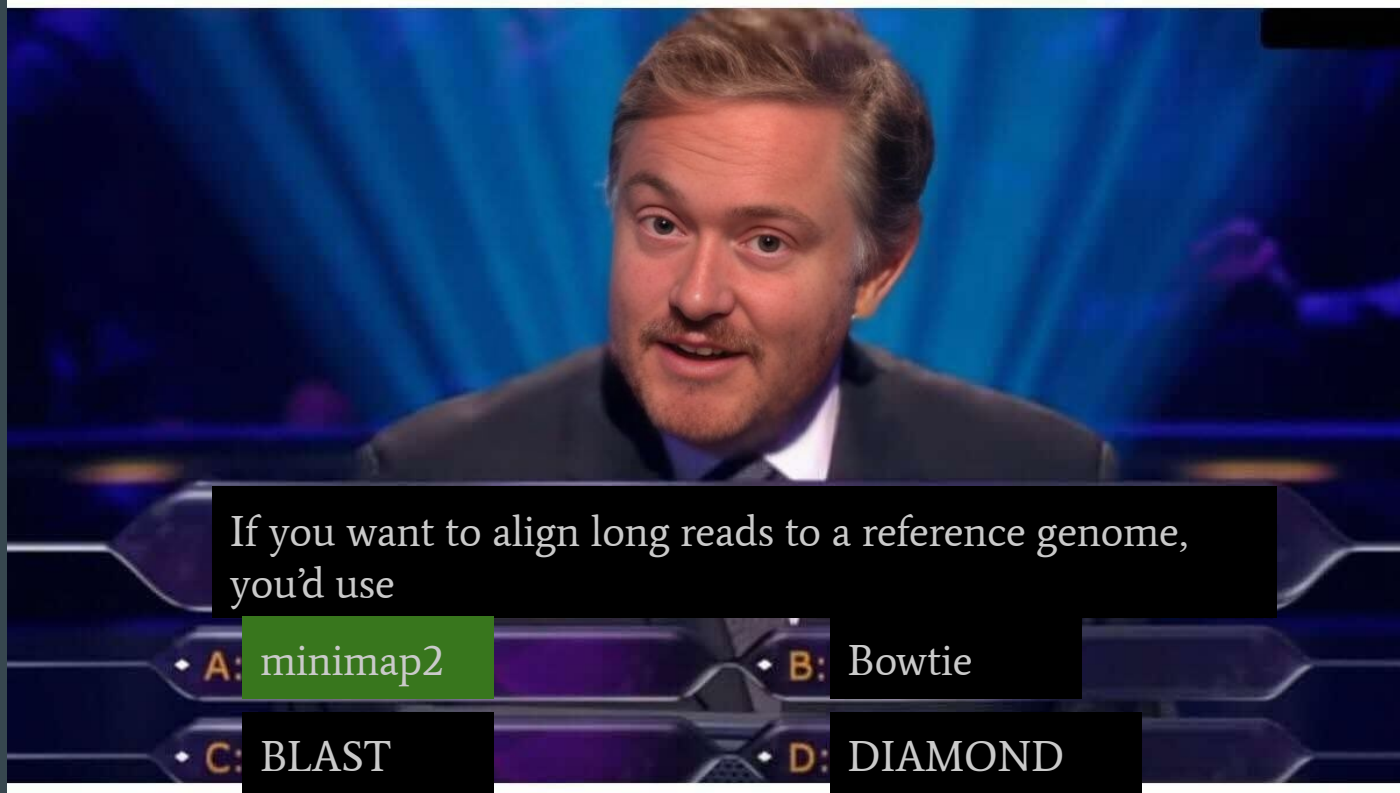
<https://github.com/soedinglab/mmseqs2/wiki#description-of-workflows>

# How DIAMOND work:

*“[..] A simple exact match criterion determines which seeds are passed on to the extension phase, in which a Smith-Waterman alignment is computed.”*

<https://www.nature.com/articles/nmeth.3176>

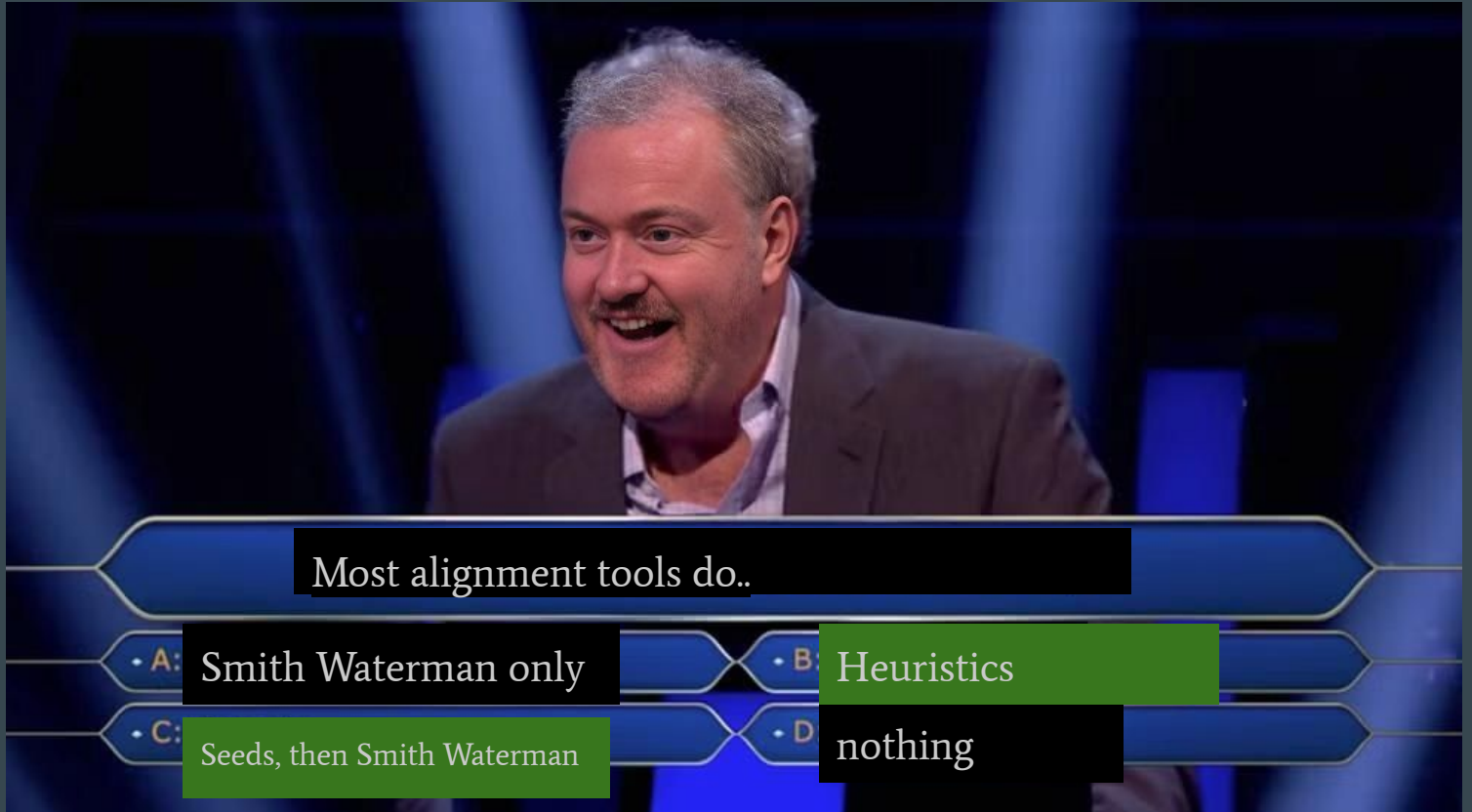
# Quiz time!

A man with a mustache, wearing a dark suit and tie, is shown from the chest up. He has a surprised or excited expression. The background is a blue, textured wall with some lights. In the foreground, there is a black overlay with white text and four options labeled A, B, C, and D. Option A is highlighted in green.

If you want to align long reads to a reference genome, you'd use

- A: minimap2
- B: Bowtie
- C: BLAST
- D: DIAMOND

# Quiz time!



Most alignment tools do..

- A: Smith Waterman only
- B: Heuristics
- C: Seeds, then Smith Waterman
- D: nothing

**Multiple, protein?**

...

# What it looks like

Input:  $n$  sequences

ACATGA

ACGTG

CATTA

Output: aligned sequences, with indels

ACATGA

AC**G**TG-

-CAT**T**A

Colors = equivalent residues (structurally or functionally)

# In practice..

Q5E940_BOVIN	-----MPREDRATWKSNYFLKIIQLDDYPKCFIVGADNVGSKOMQOIRMSLRGK-AVVLGMGKNTMMRKAIRGHLENN--PALE	76
RLA0_HUMAN	-----MPREDRATWKSNYFLKIIQLDDYPKCFIVGADNVGSKOMQOIRMSLRGK-AVVLGMGKNTMMRKAIRGHLENN--PALE	76
RLA0_MOUSE	-----MPREDRATWKSNYFLKIIQLDDYPKCFIVGADNVGSKOMQOIRMSLRGK-AVVLGMGKNTMMRKAIRGHLENN--PALE	76
RLA0_RAT	-----MPREDRATWKSNYFLKIIQLDDYPKCFIVGADNVGSKOMQOIRMSLRGK-AVVLGMGKNTMMRKAIRGHLENN--PALE	76
RLA0_CHICK	-----MPREDRATWKSNYFMKIIQLDDYPKCFVVGADNVGSKOMQOIRMSLRGK-AVVLGMGKNTMMRKAIRGHLENN--PALE	76
RLA0_RANSY	-----MPREDRATWKSNYFLKIIQLDDYPKCFIVGADNVGSKOMQOIRMSLRGK-AVVLGMGKNTMMRKAIRGHLENN--SALE	76
Q7ZUG3_BRARE	-----MPREDRATWKSNYFLKIIQLDDYPKCFIVGADNVGSKOMQOIRMSLRGK-AVVLGMGKNTMMRKAIRGHLENN--PALE	76
RLA0 ICTPU	-----MPREDRATWKSNYFLKIIQLDDYPKCFIVGADNVGSKOMQOIRMSLRGK-AVVLGMGKNTMMRKAIRGHLENN--PALE	76
RLA0_DROME	-----MVRENKAAWKAQYFVKVLEFDEFKCFIVGADNVGSKOMQOIRMSLRGK-AVVLGMGKNTMMRKAIRGHLENN--PALE	76
RLA0_DICDI	-----MSGAG-SKRKRLFTEKATKLFVYDKMIVAEADFVGS-SOLQKIRKSIRGI-GAVLMGKNTMIRKVVIRDLADSK--PELD	75
Q54LP0_DICDI	-----MSGAG-SKRKNVFTEKATKLFVYDKMIVAEADFVGS-SOLQKIRKSIRGI-GAVLMGKNTMIRKVVIRDLADSK--PELD	75
RLA0_PLAF8	-----MAKLSKQKKQMYIEKLSLIQQYSKILIVHVDNVGSKOMASVRKSLRGK-ATILMGKNTTRIRTAALKKNLQAV--PQIE	76
RLA0_SULAC	----MIGLAVTTTKKIAKWKVDEVAELTEKLTHTITIIANIEGFPADKLHEIRKKLRGK-ADIKVTKNTLNFNALKNAG----YDTK	79
RLA0_SULTO	----MRIMAVITQERKIAKWKIEEVKLEKLEKREYHTITIIANIEGFPADKLHDIRKKMRGM-AEIKVTKNTLFCIAAKNAG----LDVS	80
RLA0_SULSO	----MKRLALALKQRKVASWKLLEEVKELTELIKNSNTILIGNLEGFADKLHEIRKKLRGK-ADIKVTKNTLFCIAAKNAG----IDIE	80
RLA0_AERPE	MSVSVLVGQMYKREKPIPEWKTLMLELELEFSKRRVVFADLTGPTFVVRVRKLLWKK-YPMVAKKRIILRAMKAAGLE--LDDN	86
RLA0_PYRAE	MMLAIGKRRYVRTQYPAKVKIVSEATELLQKYPVYVFLDLHGLSLRILHEYYRYRLARY-GVIKIIKPTLFKTAFTKYVGG--IPAE	85
RLA0_METAC	----MAEERHHEHIPQWKKDEIENIKELIQSHKVFVGMVIEGILATKMKQIRRDLDKV-AVLKVSRLTLTERALNQLG----ETIP	78
RLA0_METMA	----MAEERHHEHIPQWKKDEIENIKELIQSHKVFVGMVIEGILATKMKQIRRDLDKV-AVLKVSRLTLTERALNQLG----ESIP	78
RLA0_ARCFU	----MAAVRGS--PPEYKVRAVEEIKRMISSKPVVAIVSFRNVPAGOMQKIRREFRGK-AEIKVVKNTLLERALDALG----GDYL	75
RLA0_METKA	MAVKAKGQPPSCYEKVAEWRKREVKELKELMDEVENVGLVDLEGPAPQLQETIRAKLRERDTIIRMSRLTLMRALAEKLDER--PELE	88
RLA0_METTH	-----MAHVAEWKKKEVQELHDLIKGEYVVGIANLADIPARQLQKMRQTLDSD-ALTRMSKTLISLALAKAGREL--ENVV	74
RLA0_METTL	----MITAESEHKIAPWKIEEVNKLKLELLKNGQIVALVDMMEVPARQLQKIRDKIR-GMTLLKMSRNTLIEIAIKEVAEETGNPEFA	82
RLA0_METVA	----MIDAKSEHKIAPWKIEEVNALKLELLKSNVIALIDMMEVPARQLQKIRDKIR-DQMTLLKMSRNTLIEIAIKEVAEETGNPEFA	82
RLA0_METJA	----METKVKAHVAPWKIEEVNKLKLELLKSNVIALIDMMEVPARQLQKIRDKIR-DKVKLRMSRNTLIEIAIKEVAEELNPKLA	81
RLA0_PYRAB	-----MAHVAEWKKKEVEELANLKSYPVIALVDVSSMPAYPLSQMRRLLIRENGGLLRVSRNTLIEIAIKKAAGELGKPELE	77
RLA0_PYRHO	-----MAHVAEWKKKEVEELANLKSYPVIALVDVSSMPAYPLSQMRRLLIRENGGLLRVSRNTLIEIAIKKAAGELGKPELE	77
RLA0_PYRFU	-----MAHVAEWKKKEVEELANLKSYPVIALVDVSSMPAYPLSQMRRLLIRENGGLLRVSRNTLIEIAIKKAAGELGKPELE	77
RLA0_PYRKO	-----MAHVAEWKKKEVEELANLKSYPVIALVDVAGVPAYPLSKMRDKLE-GKALLRVSRNTLIEIAIKKAAGELGKPELE	76
RLA0_HALMA	----MSAESERKTETIPEWKQEEVDVAIVMIESYESVGVVNIAGIPESRQLDMRRDLHGT-AELRVSRNTLIEIALDDVD----DGLE	79
RLA0_HALVO	----MSESEVRQTEVIPQWKREEVDLVDVIESYESVGVVGVAGIPESRQLDSMRRLHGS-AAVRMSRNTLVNRALEDVN----DGFV	79
RLA0_HALSA	----MSAEEQRTTEEVPEWKRQEVAVLVDLLETYDSVGVVNVGTGIPESKQLDMRRGLHGQ-AALRMSRNTLLVRALEEAG----DGLD	79
RLA0_THEAC	-----MKEVSOQKKELVNEITRIKASRSVAIVDTAGIRTRQIQDIRGKNGK-INLKVIKKTLFLKALENLGD----EKLS	72
RLA0_THEVO	-----MRKINPKKKEIVSELAQDITKSKAVAVDVKGVTRROMODIRAKNRDK-VKIKVYKKTLLFKALDSIND----EKLT	72

# Why do multiple alignment?

- Comparative genomics
- Phylogeny
- Protein structure prediction
- RNA structure and function
- ...



# How is a MSA scored?

“Sum-of-pairs ” (SP) score:

- 1) Fix a scoring scheme, e.g. match=1, mismatch=-1, indel=-2.
- 2) For each column, for all pairs of residues, compute score
- 3) Sum scores across columns

Column: 123456

ACATGA

AC**G**-G-

-CAG**T**A

For column 4:  $\text{score}(T,-) + \text{score}(T,G) + \text{score}(-,G) = -2 + -1 + -2 = -5$ .

For column 5:  $\text{score}(G,G) + \text{score}(G,T) + \text{score}(G,T) = 1 + -1 + -1 = -1$ .

# Optimal MSA

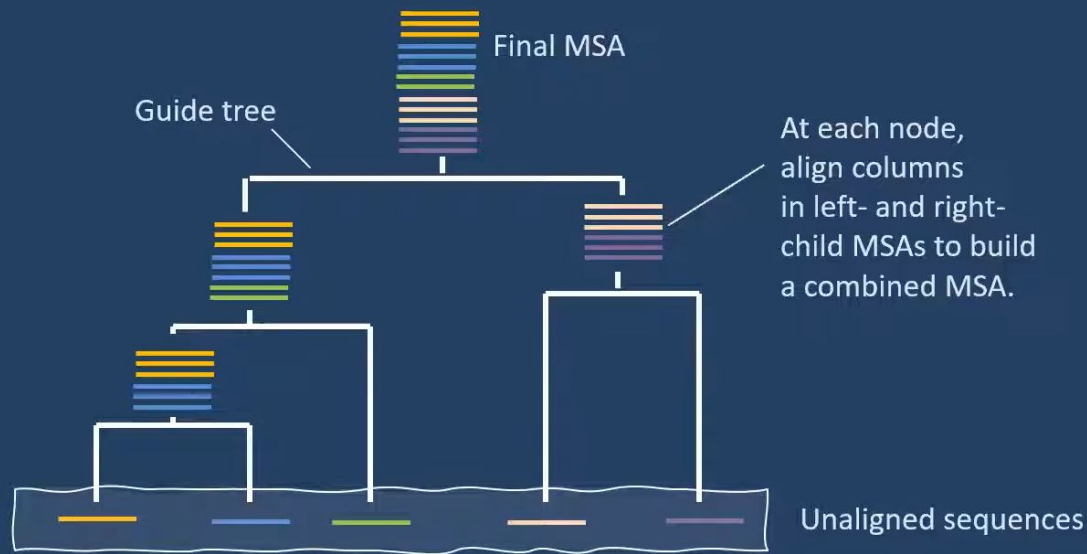
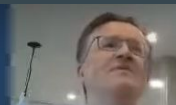
Remember Needleman-Wunsch?

Same, but with more possibilities.

So, best avoided.

# Progressive MSA

## Progressive alignment



# MSA is on another level of difficulty

## Challenging alignment

FLVRESQRNPQG-FVLSLC	HLQ---KVKHY
FIIRFSERNPGQ-FGIAYI	GVEMPARIKHY
FLLRFSESSREGAITFTWV	--E---RSQNG
FLVRDASTKMHGDYTLTLR	--K---GGNNK

FLVRESQRNPQG-FVLSLC	HLQ----KVKHY
FIIRFSERNPG-QFGIAYI	GVEMP-ARIKHY
FLLRFSESSREGAITFTWV	ERSQNGGEPD-F
FLVRDASTKMHGDYTLTLR	--K---GGNN-K

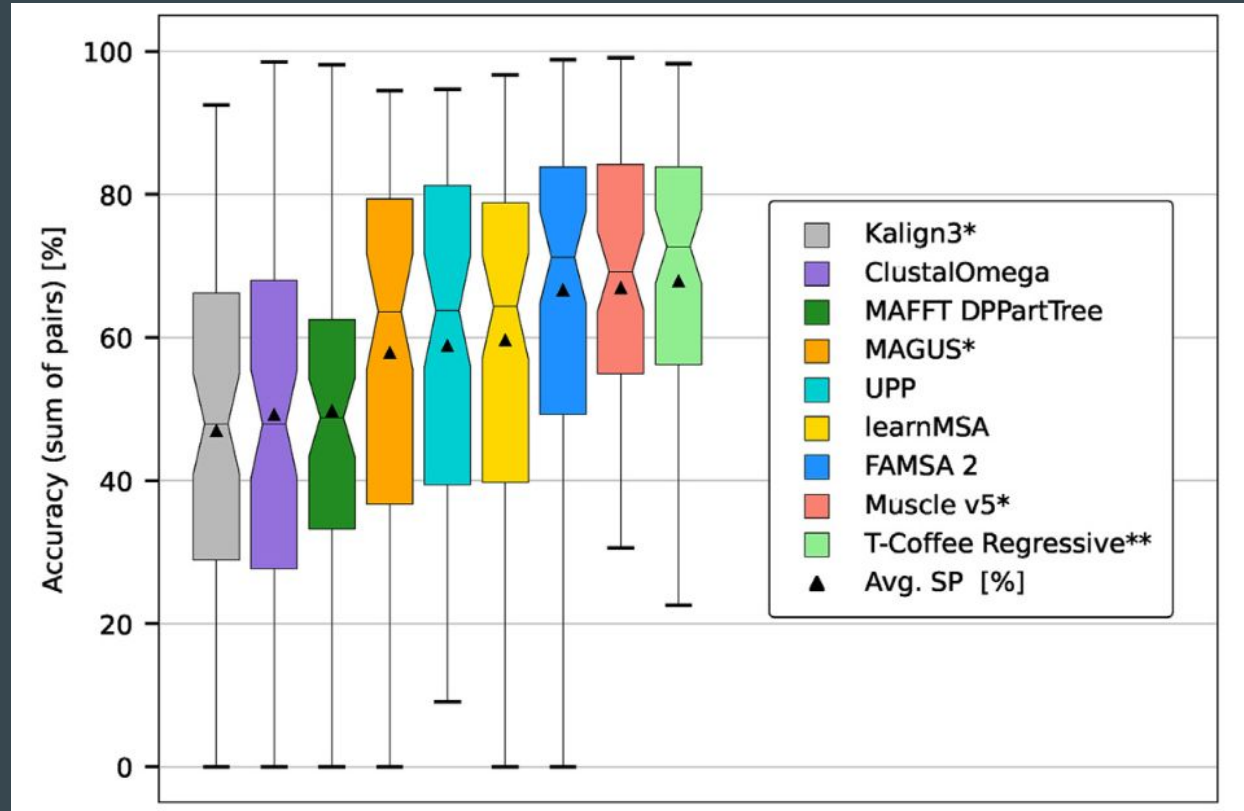
Alternative MSAs  
of same sequences

Which one is  
correct / better?

Hard / impossible  
to decide, even  
with structures

# Tools

- MUSCLE
- ClustalW
- T-Coffee
- MAFFT
- ...



<https://www.sciencedirect.com/science/article/pii/S0959440X23000519>

# Multiple, DNA

...

# What changes compared to protein MSA?

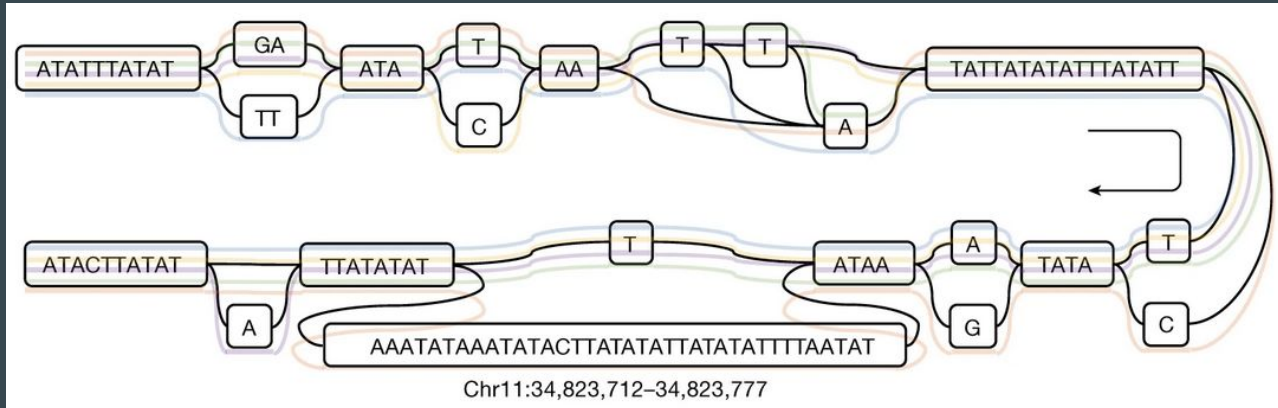
- Wayyy longer sequences
- Duplications, inversions, and translocations wreak linearity

# Tools

- SibeliaZ
- Cactus

State of the art: human genome graphs, look for pangenomics papers.

e.g. HPRC: <https://www.nature.com/articles/s41586-023-05896-x>, CPC <https://www.nature.com/articles/s41586-023-06173-7>





# 1 sequence versus a profile

...

PSSMs, HMMs

Is there enough time to present this?!

# Position Specific Scoring Matrices (PSSM) and Hidden Markov Models (HMM)

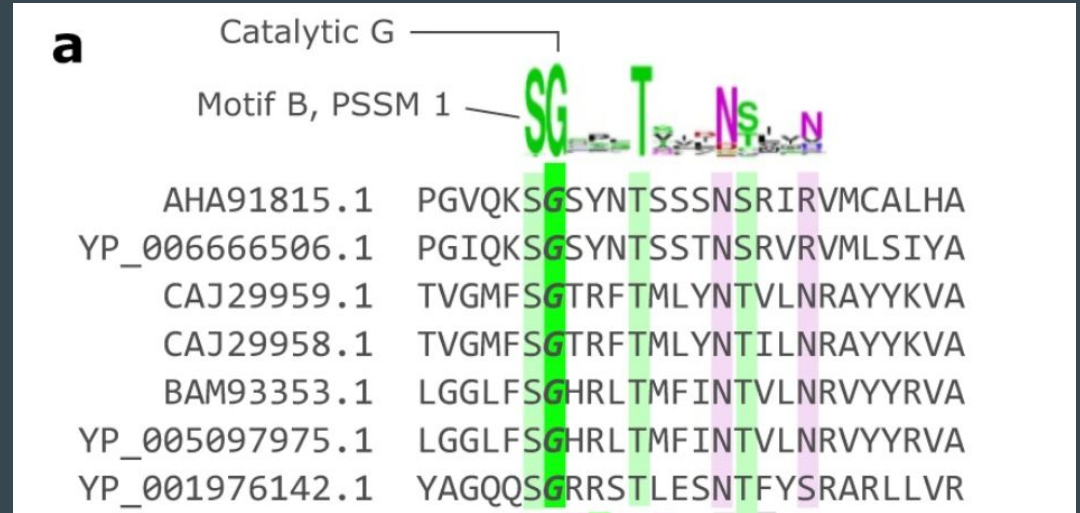
Not quite alignment, but:

*“Does this sequence belong to a particular family?”*

# PSSM

Way to represent families of sequences, **with no gaps**.

- 1) Construct MSA
- 2) Determine frequency per column

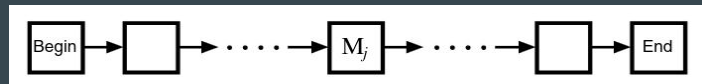


# HMM

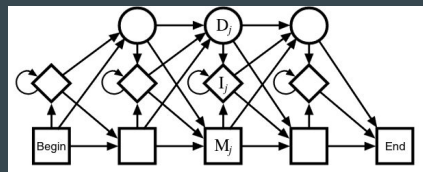
Hidden Markov Models generalize PSSMs with gaps.

Motivation: when pairwise fails

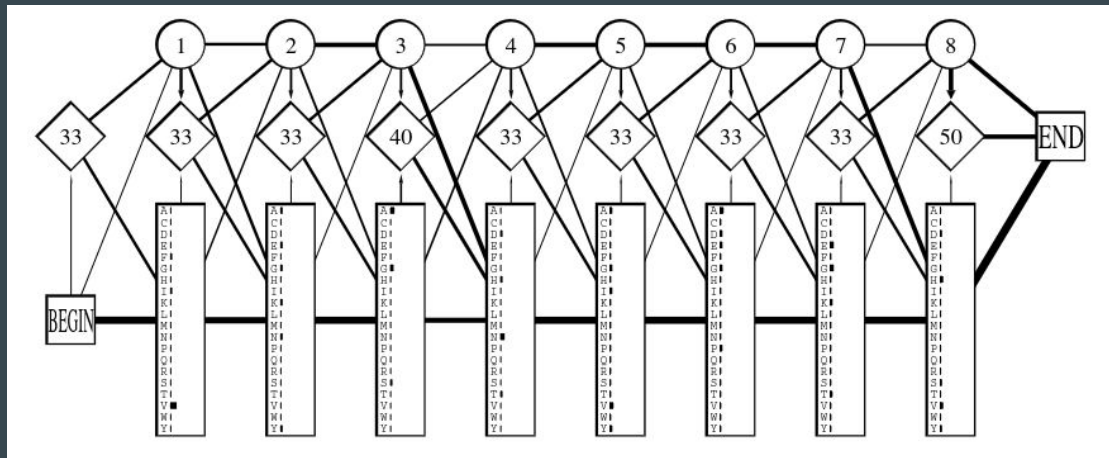
HMM of a PSSM:



Profile HMM:



HBA_HUMAN	...VGA--HAGEY...
HBB_HUMAN	...V----NVDEV...
MYG_PHYCA	...VEA--DVAGH...
GLB3_CHITP	...VKG-----D...
GLB5_PETMA	...VYS--TYETS...
LGB2_LUPLU	...FNA--NIPKH...
GLB1_GLYDI	...IAGADNGAGV...
	*** *****



# Tools

HMMer

MMseqs profile

HHblits

# Bonus: structural alignment (TM-align)

(":" denotes aligned residue pairs of  $d < 5.0$  Å)

```
MVLSEGEWQLVLHVWAKVEADVAGHGQDILIRLFKSHPETLEKFD R V K H L K T E A E M K A S E D L K K H G V T V L T A L G A I L K K K --G-HH E A E L K P L A Q S
```

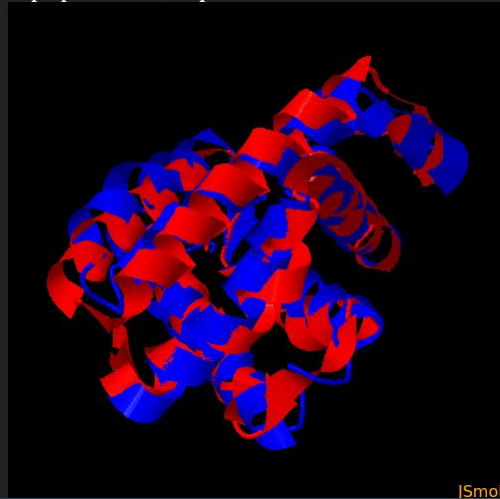
```
.....:.....:.....:
```

```
-SLSAAEADLAGKSWAPVVFANKNANGLDFLVALFEKFPDSANFFADFKGKS-VADIKASPCLRVDSSRI F T R L N E F V N N A A N A G K M S A M L S Q F A K E
```

Input: 2 PDB structures

Output: aligned residues,  
and a TM -score  
( $> 0.5$  = same fold)

Superposition of two proteins



Max TM -score:  
0.85377

# Personal take

As databases of genomes grow, alignment will both become easier and harder.

Solved:

- Human read alignment (DNA, RNA)
- High-identity to current genome databases
- Small-data HMMs

Unsolved:

- Genome-scale MSA
- Ancient DNA
- Large MSAs
- Big-data HMMs
- Sequences to peta-scale databases

# What we've seen

- Pairwise DNA alignment
  - CIGAR strings
  - Scoring
  - Needleman-Wunsch
  - Smith-Waterman
  - BLAST
  - BLAT, minimap2
- Short read mapping
  - Burrows-Wheeler transform
  - Minimizers
  - Bowtie2, BWA, minimap2, Strobealign
- Pairwise protein alignment
  - Diamond, mmseqs2
- MSA
- HMMs



Thank you for your attention!

